

CLINICAL COUNTERPOINT: Vitamin D: New Actions, New Analogs, New Therapeutic Potential*

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- I. Introduction
- II. Promising Analogs
 - A. Natural metabolism
 - B. Synthetic modifications
 - C. Importance of binding affinities
 - D. Genomic and nongenomic responses
 - E. Structure-function studies
- III. Clinical Applications
 - A. Metabolic bone disease
 - 1. Background
 - 2. Osteoporosis
 - 3. Osteopetrosis
 - 4. Hyperparathyroidism
 - 5. Chronic renal failure
 - 6. Hypophosphatemic rickets
 - B. Cancer
 - 1. Epidemiology
 - 2. Laboratory and animal studies
 - 3. Tumor production of $1,25(\text{OH})_2\text{D}$
 - C. Immune function
 - 1. Cellular effects of calcitriol
 - 2. Production of calcitriol by macrophages
 - 3. Clinical studies
 - 4. Animal studies
 - D. Hypertension
 - 1. Role of calcium
 - 2. Role of vitamin D
 - E. Diabetes mellitus
 - 1. Calcitriol regulation of insulin secretion
 - 2. Insulin regulation of calcitriol production
 - 3. Diabetes and metabolic bone disease
 - F. Psoriasis
 - 1. Targets for calcitriol action
 - 2. Clinical studies
- IV. Conclusions

I. Introduction

AN EXCITING new era has developed in the vitamin D field with the discovery of new target tissues,

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mechanisms of action, and selective analogs. As a clinical counterpoint article, this review will emphasize the potential clinical applications of these new discoveries. Many of the applications to be discussed remain untested, whereas others remain in their infancy. A major reason for optimism in predicting an expanding list of therapeutic applications for the vitamin D metabolites and analogs stems from the development of analogs of calcitriol that differ in their biological effects. In particular, analogs are now available that appear to separate the effects of calcitriol on growth and differentiation from effects on intestinal calcium absorption or bone mobilization. This apparent selectivity may reflect altered pharmacokinetic properties or may involve mechanistic differences at the cellular level as well. Regardless, the development of these drugs is likely to lead to clinical applications in which raising serum calcium need not accompany other actions. The initial portion of this review is devoted to a discussion of these analogs. The main goal of this article is to review those areas in which calcitriol and its analogs are being used in new ways and to describe potential applications that are suggested by the newly discovered actions of calcitriol. Thus, new concepts in vitamin D action have led to clinical trials of calcitriol and its analogs in the management of hyperparathyroidism and psoriasis, and trials of these drugs in certain malignancies or immunological disorders may not be far off. None of the clinical applications discussed are approved applications for these drugs. Rather, this review is intended to link the laboratory observations of the past decade with the bedside of the next decade.

In the past decade, a number of tissues have been found to contain receptors for the active vitamin D metabolite, $1,25(\text{OH})_2\text{D}$, and to respond to this hormone with a change in function. The classic target tissues, bone, kidney, and intestine, responsible for maintaining bone mineral homeostasis in response to vitamin D and its metabolites are now only part of a list that includes several dozen tissues including various elements of the hematopoietic and immune system, cardiac, skeletal, and

smooth muscle, brain, liver, breast, endothelium, skin (keratinocytes, melanocytes, and fibroblasts), and endocrine glands such as the pituitary, parathyroid gland, pancreatic islets (β -cells), adrenal cortex and medulla, thyroid, ovary, and testis. Furthermore, malignancies developing within these tissues may also contain vitamin D receptors (VDR) and be expected to respond to $1,25(\text{OH})_2\text{D}$. The responses of these tissues to $1,25(\text{OH})_2\text{D}$ are as varied as the tissues themselves. $1,25(\text{OH})_2\text{D}$ regulates hormone production and secretion including insulin from the pancreas, PRL from the pituitary, and PTH from the parathyroid gland just as it regulates cytokine production and secretion such as interleukin-2 (IL-2) from the lymphocyte and tumor necrosis factor from the monocyte. Myocardial contractility and vascular tone are modulated by $1,25(\text{OH})_2\text{D}$ as is liver regeneration. $1,25(\text{OH})_2\text{D}$ reduces the rate of proliferation of many cell lines including normal keratinocytes, fibroblasts, lymphocytes, and thymocytes as well as abnormal cells of mammary, skeletal, intestinal, lymphatic, and myeloid origin. Differentiation of numerous normal cell types including keratinocytes, lymphocytes, hematopoietic cells, intestinal epithelial cells, osteoblasts, and osteoclasts as well as abnormal cells of the same lineage is enhanced by $1,25(\text{OH})_2\text{D}$. Thus, the potential for manipulating a vast array of physiological and pathological processes with vitamin D-related compounds is enormous.

The major problem facing the clinician desiring to manipulate any one of these newly recognized actions of vitamin D is that $1,25(\text{OH})_2\text{D}$ is likely to require higher than physiological doses to be effective and will not be selective at such doses. Thus, to use $1,25(\text{OH})_2\text{D}$ to treat diabetes mellitus, control psoriasis, or modulate the growth of the tumor is to risk complications associated with hypercalcemia and hypercalciuria. Developing analogs of $1,25(\text{OH})_2\text{D}$ to improve the selectivity and confer a lower risk-benefit ratio has become a major effort by several pharmaceutical firms, and the early results look quite promising. In a summary of their experience with 228 analogs of vitamin D, Norman *et al.* (1) have suggested that different analogs bind to the VDR from different tissues with different affinities (although this may be a species rather than tissue difference), that the affinity of an analog for the VDR in cells does not necessarily parallel its affinity for the circulating vitamin D binding protein (DBP), and that the affinity of an analog for DBP or VDR does not necessarily predict its biological activity. As will be discussed in more detail below, different cells respond differently to the various analogs, and, indeed, some of these analogs are being used in clinical trials for conditions such as psoriasis without apparent risk of hypercalcemia.

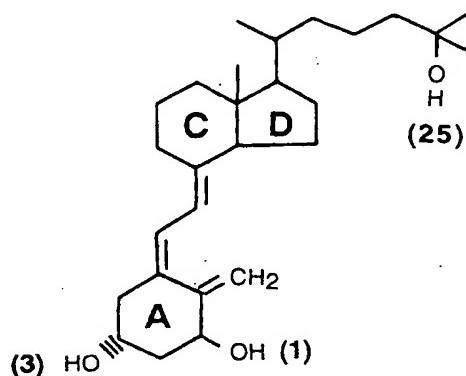
In this clinical counterpoint to Marian Walters' exten-

sive review of newly identified actions of the vitamin D endocrine system (2) I will review recent progress in the development of vitamin D analogs that show clinical promise before examining clinical conditions in which these analogs or $1,25(\text{OH})_2\text{D}$ (calcitriol) itself have been used or could conceivably be used in the near future. Most of the conditions that will be discussed are not approved indications for the use of calcitriol, and none of the analogs to be discussed have been approved for clinical use. Nevertheless, the trends are clear as we move into a new era in the therapeutic application of vitamin D metabolites and analogs. In this discussion calcitriol and $1,25(\text{OH})_2\text{D}$ will be used interchangeably.

II. Promising Analogs

A: Natural metabolism

Ever since the discovery that vitamin D required metabolism first to 25-hydroxyvitamin D (25OHD) and then to a variety of metabolites, the most important of which is $1,25(\text{OH})_2\text{D}$, considerable effort has been expended in determining the structure-function relationships of this family of vitamin D seco-steroids. The early work concerning these structure-function relationships has been reviewed previously (3, 4). The salient features are as follows. The most potent naturally occurring vitamin D seco-steroid in terms of stimulating intestinal calcium transport, mobilizing calcium from bone, raising serum calcium, and healing rickets (the "classic" actions of vitamin D) is $1,25$ -dihydroxyvitamin D ($1,25(\text{OH})_2\text{D}$). As depicted in Fig. 1, this molecule has three hydroxyl groups, two of which are in the A ring on opposite sides of the plane of the ring (3β -OH and 1α -OH) and the third in the side chain (25 OH). The 1α - and 25 OH



1,25(OH)₂D₃

FIG. 1. The structure of $1,25(\text{OH})_2\text{D}$ (CT). The three hydroxyl groups are numbered as to position as are the ring structures.

groups are critical for binding to the VDR. Removal of either OH group [as in 1α -OH D₃ or 25-hydroxyvitamin D₃ (25OHD₃)] reduces the affinity of the molecule for the VDR by approximately 3 orders of magnitude, and removal of both (as in vitamin D₃ itself) essentially eliminates binding to the VDR. In contrast, DBP has a higher affinity for 25OHD₃ than for either 1,25-dihydroxyvitamin D₃ [1,25(OH)₂D₃] or vitamin D₃ (5). Correlating with their importance for binding to the VDR, the 1α and 25 OH groups are required for full biological activity at least in terms of their classic actions in regulating calcium homeostasis. The 3β OH group is of lesser importance. Removing the 3β OH group reduces both affinity for the VDR and biological activity by less than 1 order of magnitude (6). The tricene structure linking ring A and ring C is in the cis configuration in the naturally occurring metabolites. Rotation of ring A into the transconfiguration brings the 3β -OH into a pseudo 1α position. The presence of the pseudo 1α -OH in dihydrotachysterol (DHT) accounts for its greater biological activity compared to vitamin D in nephrectomized animals or patients with chronic renal failure who have lost their capacity for 1α -hydroxylation of the vitamin D seco-steroids. DHT is an early example of a clinically useful analog. 25-Hydroxy-DHT₃ (25OHDHT₃) binds to the VDR with 10 times higher affinity than 25OHD₃ and is the presumed biologically active metabolite of DHT (7). The side chain is the site of extensive modification by the body as well as by organic chemists (as will be discussed below). The 24 position is hydroxylated by many tissues, the OH group being inserted into the R position. Although the biological importance of 24,25(OH)₂D remains in dispute, the insertion of the 24(R) OH group into 1,25(OH)₂D₃ reduces its classic biological activity and affinity for the VDR (8, 9). Although the insertion of 24 OH adjacent to 25 OH in the side chain reduces the biological activity, 24 OH can substitute for 25 OH with little loss of activity (10). In other words, for optimal activity either a 24 or a 25 OH group is needed, but not both. The C23 and C26 positions also undergo hydroxylation, metabolic changes that reduce the biological potency of the parent molecule and appear to be part of the catabolic pathway for the active vitamin D metabolites (reviewed in Ref. 11). However, evidence has been obtained suggesting that 25,26(OH)₂D₃ stimulates intestinal calcium transport without raising serum calcium (12), making this metabolite an early example of a vitamin D compound with selective biological function.

B. Synthetic modifications

Although nature can produce a large number of metabolic alterations in the vitamin D molecule, organic

chemists have vastly expanded the repertoire. A major incentive for this effort is to produce compounds with selectivity for one type of tissue or disease process that can be exploited clinically. Table 1 summarizes data from studies (4, 13-40) with a few of these analogs chosen either to illustrate certain points regarding structure-function relationships or to indicate trends in the clinical potential for these compounds. Figure 2 shows the structure of these analogs. In contrast to the earlier studies of metabolites and analogs summarized in the preceding paragraph and gathered during the 1970s, recent studies with newer analogs have taken advantage of models for the newly recognized actions of vitamin D. In particular, analogs have been sought that decrease proliferation and enhance differentiation of tumor cell lines and normal cells *in vitro* without increasing intestinal calcium absorption, serum calcium, bone resorption, or renal calcium excretion *in vivo*. Examples of tumor cell lines evaluated include HL-60 (a human promyelocytic leukemia cell line), U937 (a human promonocytic cell line), WEHI-3 (a mouse myelomonocytic leukemia cell line), and ROS 17/2.8 (a rat osteosarcoma cell line); examples of normal cells evaluated include keratinocytes, lymphocytes, and bone cells.

C. Importance of binding affinities

In interpreting the results of *in vitro* and *in vivo* studies, the importance of the different affinities of the analogs for both the cellular VDR and the serum DBP must be borne in mind. Thus, compounds such as 24,24-dihomo calcitriol (MC1147), 16 ene, 23 yne calcitriol (16ene, 23yne CT), 22 oxa calcitriol (OCT), and calcipotriol (MC903) have an affinity for the VDR that is within 1 order of magnitude of the affinity of calcitriol (CT). Yet these compounds are much less tightly bound to DBP. This produces several complicating results. *In vitro*, most cells are grown and studied in the presence of serum that contains DBP. We (41) have shown that the free CT concentration in culture media containing 10% fetal bovine serum is approximately 0.5% of the total concentration, and that the keratinocyte senses the free, not the total, concentration. These results have been confirmed by Bouillon *et al.* (13). They demonstrated that the ability of CT to inhibit the proliferation of phytohemagglutinin (PHA)-activated lymphocytes was inhibited 100-fold by the restoration of the normal concentration of DBP to DBP-depleted serum used at 10% concentration in the media for the experiment. In contrast, 24,24 dihomo CT and calcipotriol, which are much less tightly bound to DBP than CT, were proportionately much less affected by the presence of DBP. Thus, in evaluating the potency of a compound *in vitro* one must compare the free concentrations, not the total

TABLE 1. Structure-function relationships of vitamin D analogs

Compound	<i>In vivo</i>		<i>In vitro</i>			Antiproliferation			Prodifferentiation			Binding	
	ICA	SC _a	Bone	Immune	Skin	Bone	Immune	Skin	Bone	VDR	DBP	DBP	
1 α ,25(OH) ₂ D ₃	100	100	100 (13), (11) (13)	100	100	100	100	100	100	100	100	100	
Shortening 24 nor 1,25D ₃	0 (4)	0 (4)											
26,27 bis nor 1,25D ₃													
Lengthening 24 homo 1,25D ₃	100 (23, 28)	0 (23), 20-30 (28, 32)	100 (22)	200 (32)									
26 homo 1,25D ₃	100 (28)	300 (28)	100 (22)										
24,24 dihomo 1,25D ₃	1 (13)-10 (23)	0 (13)-4 (13, 23, 32)	0 (22)	3 (13, 13), 200 (32)									
24,24,24 trihomo 22,23,1,25D ₃	0 (23)	0 (23)	0 (22)										
Stabilizing 24,24F ₂ 1,25D ₃	300-1000 (35, 36)	100-300 (35, 36)		100		400-700 (37)		300		100 (14, 37)			
Other													
16 ene, 23 yne	0 (13), 3 (24, 26), 0 (11, 2 (24, 26, 39)		400 (24)-1200 (26)	100		200-1000 (13, 2)		300		45-300 (13, 14, 1)		0-0.2 (13, 16)	
1,25D ₃	53 (39)	1 (29, 30, 33)	2 (31)	100 (15)-1000		(4, 26)				(7, 24, 26)			
22 oxa 1,25D ₃	<1 (33)			(31, 40)		100 (15)-1000				10 (15)-100		0.2-0.4 (16, 40)	
Calcipotriol	<1 (13)	3 (13, 27)	10-30 (18)	30 (13), 100 (27)	100 (38)	100 (20)	100 (13, 27)	100 (13, 27)	10-100 (38)	100 (20), 1-10 (25)	60-300 (13, 18, 27)	2 (16)	

Underlined references indicate samples studied in presence of serum, generally 2 to 10%. ICA, Intestinal calcium absorption; SC_a, serum calcium.

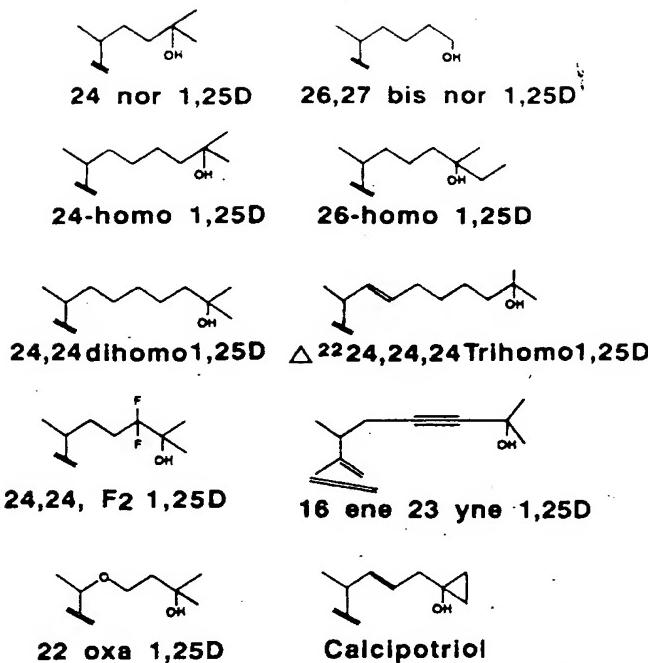


FIG. 2. Side chain modifications of CT. The analogs shown are those listed in Table 1 and discussed in the text.

concentrations. In Table 1, I have indicated which studies were performed in the presence of DBP containing serum. Nearly all *in vitro* cell culture studies have been performed in this fashion, the exceptions being the aforementioned study of human lymphocytes by Bouillon *et al.* (13), our own unpublished studies with human keratinocytes (Bikle, D., and Pillai, S., in preparation), and the study of human bone cells by Marie *et al.* (25). Except for 16ene, 23yne CT, which has a higher affinity than CT for the VDR at least in keratinocytes (14), and 24,24-difluoro-CT (24,24 F₂ CT) which may be more slowly catabolized than CT *in vitro*, much of the apparent increase in potency of the analogs relative to CT in inhibiting proliferation and stimulating differentiation *in vitro* is due to their higher free fraction in media containing serum. The different affinities of the analogs for DBP also affect the interpretation of *in vivo* studies. As observed by Dusso *et al.* (16), OCT has a shorter half-life and higher metabolic clearance rate *in vivo* than does CT. Other analogs with little affinity for DBP are likely to show similar short half-lives *in vivo*. The rapid clearance of these analogs *in vivo* may account for at least part of their selective effects. For example, raising serum calcium levels through changes in intestinal calcium transport and bone resorption where new protein synthesis may be required (42, 43) conceivably requires a more sustained level of circulating CT than inhibition of PTH synthesis or secretion (44, 45). Thus, the ability of

OCT to inhibit PTH secretion *in vivo* without raising serum calcium levels (30) could be due to this mechanism. Furthermore, these pharmacokinetic considerations may explain why CT appears to have a greater effect on bone resorption *in vitro* (18) than would be indicated by its limited effect on serum calcium levels *in vivo* (13, 27).

D. Genomic and nongenomic responses

Although differences in the relative affinities of DBP and VDR for the various analogs compared to CT can explain part of the selectivity observed for some of the analogs, differences in the manner in which different cells respond to CT and its analogs must also be considered. Not all vitamin D-regulated events require the interaction of the VDR-hormone complex with the genome to induce or inhibit new protein synthesis. This is illustrated by the cycloheximide-insensitive vitamin D-induced changes in calcium transport across the intestinal brush border (42, 46-48) and the vitamin D-induced rapid changes in intracellular calcium and/or phosphoinositide metabolism in cells from the intestine (49), liver (50), bone (51), and skin (52). This point becomes relevant to understanding the observation by Farach-Carson *et al.* (17) that the rank order of effectiveness for analogs in increasing calcium influx into ROS 17/2,8 cells does not correlate with the rank order of the affinity of the analogs for the VDR. Thus, it appears that analogs may differ in their ability to influence genomic and nongenomic mechanisms, and that the relative importance of genomic and nongenomic mechanisms in responding to the vitamin D metabolite or analog will contribute to the selectivity of that molecule.

E. Structure-function studies

A striking feature of the analogs listed in Table 1 is the profound influence caused by modest changes in the side chain. Shortening the side chain by one carbon (e.g. 24 nor CT) essentially eliminates its potency *in vivo* (4) and reduces its effectiveness *in vitro* by 1 order of magnitude (34). Removing two or more carbons from the side chain (e.g. 26, 27 bis nor CT) reduces its *in vitro* activity another order of magnitude (34). In contrast, lengthening the side chain by one carbon (e.g. 24 or 26 homo CT) preserves *in vitro* activity and the ability to stimulate intestinal calcium transport (23, 28, 32, 34). However, bone mobilization as assessed *in vivo* is less stimulated by 24 homo CT and more stimulated by 26 homo CT than would be expected from the comparable abilities of these analogs to stimulate bone resorption *in vitro* (22, 23, 28, 32). Addition of two carbons to the side chain (e.g. 24, 24 dihomo CT or MC1147) maintains the ability of this analog to inhibit lymphocyte proliferation and stimulate HL60 and U937 differentiation *in vitro* (13, 23,

32) while decreasing its ability to stimulate bone resorption *in vitro* (22) or increase intestinal calcium transport and serum calcium *in vivo* (13, 23). Adding a third carbon to the side chain (e.g. 24, 24, 24 trihomo 22ene CT) essentially abolishes *in vivo* activity with only a modest reduction in the ability to stimulate HL60 differentiation (22, 23). Substituting fluoride for hydrogen in those sites of the CT molecule that undergo further metabolism (e.g. C24, C26, C27) retards metabolism at those sites and is expected to increase the biological half-life of the molecule, although this has not been conclusively demonstrated. As exemplified by 24,24 F₂ CT, the fluoride substitution increases the *in vitro* and *in vivo* effects of CT without altering its ability to bind to the VDR (14, 35-37). This analog may be useful for conditions in which a sustained CT effect is desired.

In contrast to 24,24 F₂ CT, but similar to 24,24 dihomo CT, the remaining three analogs listed in Table 1, namely 16ene, 23yne CT, OCT, and calcipotriol, all have limited hypercalcemic effects *in vivo* (13, 24, 26, 27, 29, 30, 33, 39), yet are potent analogs in terms of their antiproliferative and prodifferentiating effects *in vitro* (13-15, 19-21, 24, 26, 27, 31, 37-40). Although some of the difference between *in vivo* and *in vitro* effects may reflect the rapid clearance of these analogs *in vivo*, this is not the total explanation. Abe *et al.* (31) noted that OCT was several orders of magnitude less potent than CT in stimulating bone resorption *in vitro*, while approximately 1 order of magnitude more potent than CT in inhibiting proliferation and stimulating differentiation of WEHI-3 cells. Furthermore, OCT appears to be 2 orders of magnitude more potent than CT in stimulating the immune response of mice to sheep erythrocytes *in vivo* (29), and equivalent to CT in suppressing PTH secretion when administered *in vivo* (30) despite its lack of effect on serum calcium levels. An important potential difference in the mechanisms of action between OCT and CT is that OCT failed to raise the intracellular free calcium levels (Cai) in HL60 cells or enhance the fMLP-induced Cai spike and superoxide generation. Yet OCT had effects comparable to CT on proliferation and differentiation of this cell line (15). Similarly, the CT-induced increase in Cai in ROS 17/2.8 cells could not be reproduced by OCT (51). Unlike the slow response of Cai to CT in HL60 cells, the response of Cai to CT in ROS cells is acute, suggesting that the mechanisms differ between the two cell lines, yet OCT apparently fails to activate the responsible mechanism in either case. It remains to be determined whether other analogs will share this potentially important difference in mechanism of action between CT and OCT on Cai, whether this difference will be found in all target cells, and whether this difference contributes to the selectivity of OCT for the parathyroid gland and the immune system *in vivo*.

However, such differences in fundamental mechanisms of action and the pharmacokinetic differences related to differential binding to DBP and VDR indicate that vitamin D analogs can be made with selective biological properties that can be exploited therapeutically. Several of these are already in clinical trials as will be described below.

III. Clinical Applications

The potential new clinical applications that will be discussed in this section are listed in Table 2. For some, such as the use of CT (or its analogs) in the management of osteoporosis, hyperparathyroidism accompanying renal failure, or psoriasis, existing data from clinical trials are quite promising. For others, such as the treatment of immune disorders, malignancy, hypertension, or diabetes mellitus, the use of CT or its analogs appears more distant.

The newly discovered actions of CT relevant to its new therapeutic potential can be considered of two sorts: 1) modulation of hormone and cytokine production and secretion, and 2) regulation of proliferation and differentiation. CT may exert its influence on cells by actions in one or both categories. An attempt to illustrate these points is shown in Figs. 3 and 4. In Fig. 3, CT is depicted as having a positive effect on insulin secretion by the B cell of the pancreas but a negative effect on PTH secretion from the parathyroid gland. In turn, both insulin and PTH stimulate CT production by the kidney. Such experiments lead to the possible role of CT in the management of diabetes mellitus and hyperparathyroidism. In Fig. 4 CT is depicted as having an antiproliferative effect on tumor cells and basal cells of the epidermis while promoting their differentiation as well as the differentiation of the bone cell precursors for both osteoblasts and osteoclasts. Most likely these effects on proliferation and differentiation involve regulation of, or at least interaction with, a number of cytokines (and hor-

mones) in the various tissues that participate in the control of proliferation and differentiation of these tissues. Such actions suggest the usefulness of CT or its analogs in the treatment of osteoporosis, osteopetrosis, cancer, immune disorders, and psoriasis. These two recurring themes, the ability of CT to regulate hormone and cytokine secretion and its potential to modulate growth and differentiation of its "nonclassical" target tissues, underlie the development of CT and its analogs for new clinical indications.

A. Metabolic bone disease

1. Background. Although the use of vitamin D and its metabolites for the management of certain metabolic bone diseases is well established, the new understanding of the role of CT in the differentiation of osteoclasts and osteoblasts offers a rationale for the use of CT and its analogs in the management of osteoporosis and osteopetrosis. Furthermore, the discovery of the VDR in parathyroid tissue and the elucidation of its role in regulating PTH synthesis have led to a new approach in the management of primary and secondary hyperparathyroidism. In the treatment of these different conditions, hypercalcemia and/or hypercalciuria frequently limit the amount of CT that can be employed, and suboptimal doses may be required and/or dietary calcium may need to be restricted. For this reason, CT analogs with less potential for hypercalcemia may play a more important role in these clinical applications.

The amount of bone in the skeleton is controlled by the balance of bone formation and bone resorption that are mediated by osteoblasts and osteoclasts, respectively. The osteoblast contains receptors for CT; the osteoclast does not (53). Osteoblasts are responsible for bone formation, but the role of vitamin D in this process is not clear. In cultured rat or mouse calvaria and the osteoblast-like cells derived from them, CT inhibits collagen synthesis and alkaline phosphatase at concentrations

TABLE 2. Potential new clinical applications for CT and its analogs

Disease	Postulated actions
Osteoporosis	Increase bone mineral absorption from gut Enhance osteoblast differentiation and function
Osteopetrosis	Enhance osteoclast differentiation
Hyperparathyroidism	Suppress PTH synthesis
Cancer	Decrease proliferation, enhance differentiation
Immune disorders	Enhance suppressed activity in vitamin D deficiency
Hypertension	Reduce inflammatory response of activated cells
Diabetes mellitus	Reduce calcium accumulation by vascular smooth muscle Increase insulin secretion Correct decreased calcitriol production in insulin deficiency
Psoriasis	Decrease inflammatory component Decrease proliferation, enhance differentiation of epidermis

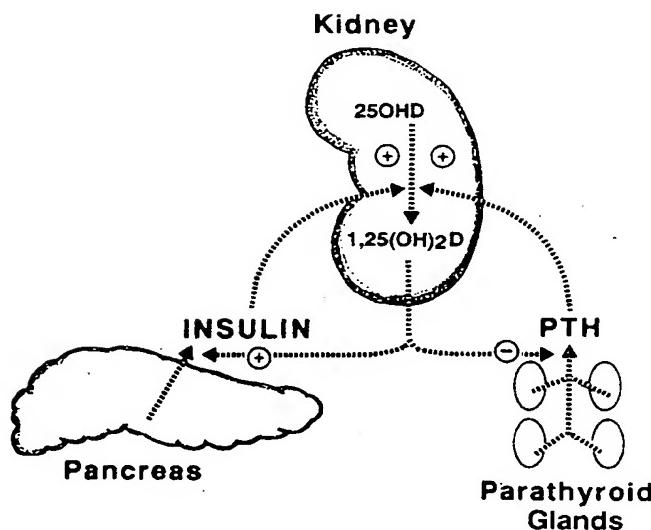


FIG. 3. Endocrine interactions of CT. This figure illustrates the rationale for the potential usefulness of CT in the management of diabetes mellitus and hyperparathyroidism. CT stimulates insulin secretion but inhibits PTH secretion. Both PTH and insulin increase CT production. In diabetes mellitus acute insulin deficiency may result in decreased CT production. Conversely, insulin secretion may be blunted in vitamin D deficiency and/or enhanced by CT supplementation. Hyperparathyroidism developing in patients with renal failure (and depressed CT production) responds to CT supplementation. Primary hyperparathyroidism may also be amenable to treatment with CT analogs that inhibit PTH secretion but do not aggravate the hypercalcemia.

comparable to those required to stimulate bone resorption (54–57). In apparently less differentiated cells, such as the mouse osteoblastic cell line MC3T3-E1 and various human osteoblastic cell lines, CT stimulates both collagen synthesis and alkaline phosphatase (19, 58). This seeming paradox may be explicable by the ability of CT to promote the differentiation of the osteoblast (59), pushing osteoblast precursors to a more mature phenotype with higher bone-forming capabilities (reflected by increased collagen synthesis and alkaline phosphatase) while inhibiting these functions in the mature cell. This may be part of the mechanism by which CT stimulates bone formation. However, the ability of calcium and phosphate supplementation alone to reverse vitamin D-deficient rickets (60) or rickets due to a dysfunctional VDR [hereditary vitamin D-resistant rickets or vitamin D-dependent rickets type II (VDDR II)] (61, 62) suggests that the direct effect of CT on the osteoblast is not essential for bone formation. The differentiated osteoblast is required for 1,25(OH)₂D-stimulated bone resorption acting as the transducer for CT to activate the osteoclast presumably by the release of cytokines that can act on the osteoclast (63). Osteoclasts do not have the VDR and do not respond to CT directly (63).

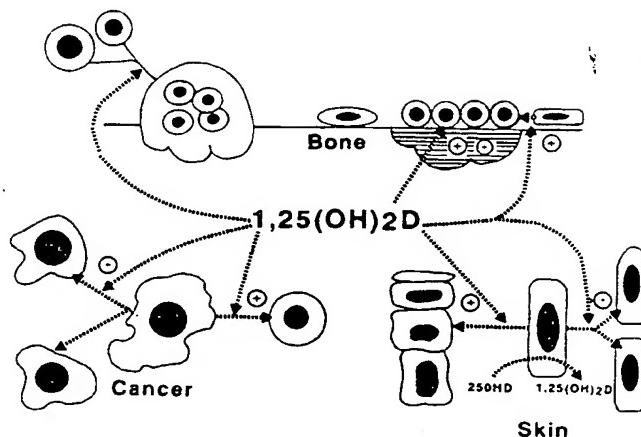


FIG. 4. Antiproliferative, prodifferentiating effects of CT. This figure illustrates the potential usefulness of CT in the management of selected bone disorders, malignancies, and hyperproliferative skin disorders. By promoting the differentiation of osteoblasts and osteoclasts, CT may be useful in the management of osteoporosis and osteopetrosis. Malignancies of cells containing the VDR may respond to both the antiproliferative and prodifferentiating effects of CT and its analogs. Psoriasis is already being treated successfully with CT and its analogs, with a decrease in the hyperproliferation and a change to a more normal appearing (*i.e.* better differentiated) skin. Note that in the epidermis, CT may have an autocrine or paracrine function because of the ability of keratinocytes to make this potent metabolite.

However, as for osteoclasts, CT is an important regulator of osteoclast differentiation, promoting their formation from hematogenous precursors that do contain the VDR (64). Thus, CT is likely to play an important role in bone remodeling not only by its ability to provide adequate calcium and phosphate for mineralization but by influencing cytokine secretion by osteoblasts and differentiation of bone cell precursors.

2. Osteoporosis. Numerous studies attempting to treat postmenopausal osteoporosis with vitamin D and its metabolites have been performed, and the results have been mixed. Three recent studies (65–67) using the same protocol in which CT was administered over a 2-yr period and the dose adjusted to avoid hypercalcemia reported somewhat different results. The study by Ott and Chestnut (66) used the lowest mean dose of the three studies and showed no effect on histomorphometric measurements or on cortical bone density measured by single or dual photon absorptiometry techniques. Trabecular bone density measured by quantitative computed tomography of the spine appeared to increase, although the number of patients serially studied by quantitative computed tomography was small. In contrast, the studies by Aloia *et al.* (65) and Gallagher and Goldgar (67) showed significant increases in spinal bone density and total body calcium compared to placebo-treated controls. None of these three studies showed a significant decrease in frac-

ture rate, and hypercalcemia and hypercalciuria complicated treatment in some of the subjects. The considerably larger 3-yr study by Tilyard *et al.* (68) evaluated 314 women treated with a twice daily dose of 0.25 µg CT compared to 308 women treated with 1 g calcium supplementation. They observed a 70% reduction in the incidence of new fractures after the first year of treatment with only two cases of hypercalcemia and one case of decreasing renal function. Likewise, an up to 8 yr study by Caniggia *et al.* (69) of 270 postmenopausal osteoporotic women treated with 1 µg CT/day without calcium supplementation showed a 75% reduction in fractures after the first year. This was accomplished without an increase in serum calcium or deterioration of renal function. However, the subjects treated with CT in these studies did have a significant increase in urinary calcium excretion. Similar results have been obtained with the CT analog, 1 α -hydroxyvitamin D (1 α OHD) (70, 71). Conceivably, analogs with less potential for increasing serum calcium relative to their effects on bone could provide a safer alternative to CT in the management of osteoporosis allowing for larger doses and less intense monitoring.

3. *Osteopetrosis.* Osteopetrosis is caused by a failure to resorb bone and is due to impaired osteoclast function most likely of several etiologies including impaired cytokine production (72, 73). Bone marrow transplant may be curative (74), attesting to the hematogenous origin of the osteoclast. In a recent evaluation of 16 infants with malignant osteopetrosis, Cournot *et al.* (75) found that many had low or low normal serum calcium and phosphate levels accompanied by increased alkaline phosphatase, PTH, and 1,25(OH)₂D levels suggesting resistance to the bone-resorbing effects of these hormones. Histological observations of the bone from six subjects revealed abundant osteoclasts in five but none in one, whereas osteoblasts were reduced in all. Although it is not clear to what extent CT can correct the abnormal function of these bone cells, CT has been used in very high doses to enhance bone resorption *in vivo* and *in vitro* with monocytes from one such patient (76). In this latter study a low calcium diet was employed to mitigate the hypercalcemia.

4. *Hyperparathyroidism.* PTH synthesis and secretion are inhibited by both calcium and CT (77). The parathyroid gland contains VDR (78, 79), which are reduced in uremia (80, 81), potentially making the parathyroid gland less sensitive to CT. Similarly, the response of the parathyroid gland to calcium is blunted in uremia, but the mechanism is unclear (82). CT inhibits PTH secretion by inhibiting its synthesis at the level of gene transcription (44, 83), and a vitamin D response element has been identified in the 5'-flanking region of the PTH gene (84).

The response of PTH secretion to CT requires hours. Unlike CT, calcium exerts a direct effect on PTH secretion, an effect seen rapidly and thought to be mediated by a calcium response element in or near the plasma membrane (85). However, calcium also reduces the messenger RNA levels for prepro-PTH indicating that it also exerts an effect on synthesis either at the level of transcription or message stability (86). Thus, nonhypercalcemic analogs of CT could be useful in the management of this condition.

5. *Chronic renal failure.* Secondary hyperparathyroidism complicates chronic renal failure, often occurring before serum calcium levels fall but generally in association with decreased 1,25(OH)₂D levels (87). It has long been appreciated that oral CT administration to patients with renal failure raises their serum calcium level, reduces their PTH secretion, and improves their clinical condition (88, 89). At least part of this effect is due to the increase in intestinal calcium absorption with CT therapy. Unfortunately, many patients with severe hyperparathyroidism are quite sensitive to CT in that they develop hypercalcemia with doses that have little impact on their PTH levels. Slatopolsky *et al.* (90) observed that the iv administration of CT reduced PTH levels more effectively and with less increment in serum calcium than oral administration. Presumably this is due to the higher levels of CT that can be achieved systemically with less exposure of the intestinal epithelium when the iv route is used. A subsequent study by Andress *et al.* (91) indicated that iv administration of CT could be used to correct the biochemical and skeletal abnormalities of secondary hyperparathyroidism in patients who could not be adequately managed with oral CT. Delmez *et al.* (92) confirmed the observation that iv CT could reduce PTH levels without raising serum calcium, and by manipulating the serum calcium with calcium infusions or low calcium dialysate baths showed that CT increased the sensitivity of the parathyroid gland to inhibition by calcium. That is, CT reset the set point to a lower calcium concentration. Similar data were obtained by Dunlay *et al.* (93). Since 1 α OHD requires 25 hydroxylation (presumably only in the liver in humans) to be active, it and the nonhypercalcemic analogs under recent development may provide oral alternatives to iv CT in the management of secondary hyperparathyroidism. These studies indicate that the secondary hyperparathyroidism of chronic renal failure will be one of the first new indications for treatment with CT or one of its analogs.

6. *Hypophosphatemic rickets.* Patients with X-linked hypophosphatemic rickets have an abnormality not only in phosphate handling but in 1,25(OH)₂D production (94-97). Their calcium levels tend to be in the low normal range. When treated with vitamin D and phosphate,

hyperparathyroidism often develops, although serum calcium levels remain normal (98, 99). This may be due to further reduction in $1,25(\text{OH})_2\text{D}$ production as a result of the phosphate therapy. CT more effectively heals the bone disease and affords better control of PTH than does vitamin D (97, 99). However, hyperparathyroidism can persist, although its impact on bone as these children grow into adults is not clear. Whether more aggressive treatment with CT or one of its nonhypercalcemic analogs should be initiated in patients with persistent hyperparathyroidism remains uncertain.

Although iv administration of CT represents an important advance in the treatment of secondary hyperparathyroidism complicating the management of renal failure and has been used successfully in the treatment of primary hyperparathyroidism (100), patients treated chronically in this fashion do develop increased serum levels of calcium and phosphate that may limit treatment (89). Therefore, nonhypercalcemic analogs are being evaluated. Brown *et al.* (30) showed that OCT, like CT, inhibited PTH secretion and reduced its gene transcription *in vitro* without raising serum calcium *in vivo*. As yet no clinical trials with OCT or the other nonhypercalcemic analogs have been performed to demonstrate their ability to inhibit PTH secretion *in vivo*.

B. Cancer

1. *Epidemiology.* Garland *et al.* have recently reviewed evidence correlating calcium and vitamin D with colon (101) and breast (102) cancer. Of 15 cancers evaluated, only these two showed a negative correlation between cancer incidence and the ambient UV light intensity when data from 87 locations throughout the United States were compiled. UV light exposure is suggested as a measure of cutaneous vitamin D production. In a large prospective study, these investigators noted a negative correlation between dietary calcium, vitamin D, serum 25OHD, and the incidence of colon cancer (103, 104). Similar data have been obtained at least for dietary calcium and colon cancer by others (105-107), although the correlation between vitamin D deficiency and breast cancer has been challenged (108). Schwartz and Hulka (109) have suggested that mortality from prostate cancer might also be linked to vitamin D because of a geographic relationship between mortality from this malignancy and UV light intensity similar to that for breast and colon cancer.

2. *Laboratory and animal studies.* Eisman *et al.* (110) detected VDR in breast cancer lines more than a decade ago, and the list has rapidly expanded to include a wide variety of malignancies from lung, cervix, bone, skin, colon, lymphatic, and hematopoietic tissue (111). In general, $1,25(\text{OH})_2\text{D}$ inhibits the proliferation and stimu-

lates the differentiation of these cell lines *in vitro* (112-121). Furthermore, vitamin D, $1,25(\text{OH})_2\text{D}$, or its analog $1\alpha\text{-OHD}$ have been shown to decrease tumor size, number, or lethality when given *in vivo* to animals in which the tumors were chemically induced (122-125) or grafted (126-128). Although a dose-response relationship between the vitamin D compound and its anticancer effect has been demonstrated *in vivo* and *in vitro*, the dose *in vivo* is restricted by toxicity. Thus, optimally effective doses in terms of preventing tumor growth increase mortality presumably by inducing hypercalcemia (125). Similar constraints limit the use of CT in the treatment of malignancies in humans (129), and as of yet no compelling study has shown its usefulness for this purpose in humans. However, OCT has recently been shown to inhibit the proliferation of human breast cells *in vitro* at doses 10-100 times less than CT and *in vivo* in mice without causing hypercalcemia (130). Likewise, 16ene, 23yne CT has been shown in mice to be more effective than comparable doses of CT in increasing survival after the injection of leukemic cells and does so without inducing hypercalcemia (131). Thus, the availability of the nonhypercalcemic analogs of CT should permit a reexploration of the role of vitamin D in the management of malignant disease.

3. *Tumor production of $1,25(\text{OH})_2\text{D}$.* Most malignancies cause hypercalcemia either by direct effects on bone (through elaboration of cytokines that induce bone resorption) or by elaborating PTH-related peptide. However, in a small number of Hodgkin and non-Hodgkin lymphomas (which except for HTLV-1 T-cell leukemia/lymphoma are seldom associated with hypercalcemia), the hypercalcemia appeared to be due to increased CT synthesis by the tumor (132-142). In one case (142) this was confirmed *in vitro*. Chemotherapy including glucocorticoids is used to treat these tumors, and successful treatment corrects both the elevated calcium and CT levels. In one report (140), recurrence of the Hodgkins disease was associated with recurrence of hypercalcemia and increased CT levels. However, it has not been established that production of $1,25(\text{OH})_2\text{D}$ by the lymphoma correlates with degree of differentiation or prognosis. One possibility is that as in sarcoidosis and other granulomatous diseases, the $1,25(\text{OH})_2\text{D}$ production occurs in abnormal macrophages within the tumor which are not subject to normal feedback inhibition or are bathed in cytokines such as interferon- γ (IFN- γ) which promote $1,25(\text{OH})_2\text{D}$ production. This aspect will be discussed in the section dealing with vitamin D and the immune system.

C. Immune function

1. *Cellular effects of CT.* Substantial evidence is accumulating that CT plays an important modulatory role in

the immune system (Fig. 5). Peripheral blood mononuclear cells acquire VDR when they are activated *in vitro* by agents such as PHA (143, 144). Activation of such cells leads to proliferation and elaboration of a variety of cytokines. CT inhibits proliferation (at the level of G1) and the production of IL-2, IFN- γ , and granulocyte macrophage colony stimulating factor by PHA-activated peripheral blood mononuclear cells (145-147). Studies on the effects of CT on interleukin-1 (IL-1) and tumor necrosis factor production have shown both stimulation and inhibition (148-151). CT stimulates H₂O₂ production in macrophages (152), monocyte adherence, and through the induction of heat shock proteins may protect the cell during the febrile response (153). *In vitro*, immunoglobulin production is depressed by CT (144, 154, 155), an effect that appears to be mediated by an inhibitory action on the helper function of T cells rather than a direct effect on B cells (144). Although the effects of CT on the immune functions of T cells are predominantly inhibitory when the cells are activated by PHA, CT is less inhibitory and may even be stimulatory when the cells are activated by phorbol esters or the anti-T3 monoclonal antibody OKT3 (149). Thus, the mechanism by which the immune system is activated could determine the degree or even the direction of immunomodulation by CT.

2. Production of CT by macrophages. The effects of CT on these immune functions may represent a paracrine or autocrine action. Activated (as by IFN- γ or lipopolysaccharide) normal macrophages make CT (156, 157), as do macrophages from granulomatous diseases such as sarcoidosis (158, 159) and tuberculosis (160, 161). Conceivably, the increased production of CT in such disease

states serves a protective function, but it may be sufficiently extensive that hypercalcemia ensues.

3. Clinical studies. Although much of our knowledge about CT-regulated immune function stems from *in vitro* studies, immune dysfunction in vitamin D-deficient or -resistant states has been observed indicating the clinical importance of these *in vitro* observations. Nutritional vitamin D deficiency is associated with increased risk of infection, and the neutrophils from such children appear to have abnormal motility and phagocytic ability (162, 163). Vitamin D deficiency has also been reported to predispose to disseminated tuberculosis (164). Toss and Symreng (165) reported a correlation between low 25OHD levels and anergy to skin testing in 63 elderly subjects; vitamin D treatment of five anergic subjects with low 25OHD levels normalized both the skin reactivity and the 25OHD level. Patients with chronic renal failure requiring hemodialysis were found to have reduced lymphocyte responsiveness to the proliferative stimulus of lectins, which was normalized by 4 weeks of 1 α OHD administration (166). Walka *et al.* (167) described a patient with VDDR II who had myelofibrosis and recurrent septicemia, and who on further investigation had abnormal neutrophil chemotaxis and an inability to produce immunoglobulin G antibodies to lipid A after each episode of septicemia. Calcium infusions corrected the myelofibrosis and the rickets but not the immunological abnormalities. Etzioni *et al.* (168) described five patients with VDDR II, four of whom had reduced white cell phagocytosis of *Candida albicans* and all of whom had decreased intracellular killing of this organism. Calcium infusion *in vivo* did not correct these abnormalities measured *in vitro*, although the calcium ionophore ionomycin corrected the *in vitro* defect in cell killing suggesting that CT may stimulate cell killing, at least in part, by raising intracellular calcium. This is of interest since the analog OCT does not raise intracellular calcium to the same degree as CT at least in HL-60 cells (49). Kitajima *et al.* (169) reported six cases with hypophosphatemic vitamin D-resistant rickets who had frequent infections (colds, cystitis, bronchitis, and pneumonia) that appeared to improve after 1 α OHD treatment. These patients had decreased numbers of natural killer cells and increased levels of leukocyte adenosine deaminase that normalized with treatment. On the other hand, a hyperfunctioning immune system as in chronic inflammatory conditions might also be treated with CT in vitamin D-replete individuals as suggested by the reduction in joint pain observed in seven of ten patients with psoriatic arthritis treated with CT (170).

4. Animal studies. Animal studies support these clinical observations and suggest a role for CT or its analogs in the management of immune disorders. Stroder (171)

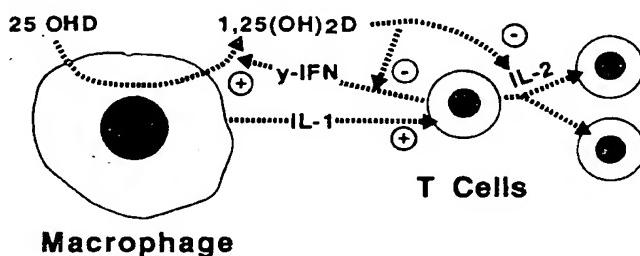


FIG. 5. Selected effects of CT on immune function. CT is made by activated macrophages and could serve to suppress the proliferation of lymphocytes by direct inhibition of the cell cycle as well as by inhibition of IL-2 secretion. Interferon- γ stimulates CT production by the macrophage, and its secretion by the lymphocyte is in turn inhibited by CT. Not shown here is that T cell activation of B cell production of immunoglobulins is also inhibited. These suppressive effects of CT on the immune system suggest that CT or its analogs may be useful in the management of inflammatory states including autoimmune disorders. However, vitamin D deficiency has been associated with decreased resistance to certain infections suggesting that CT has more than an immunosuppressive role.

reported a reduction in antibody formation in ricketic rats challenged with diphtheria toxin or Sendai virus, a more fulminant spread of pneumococcal and *Klebsiella* infection when ricketic rats were inoculated with these organisms, and a decrease in antibody forming cells in the spleen from ricketic rats. Bar-Shavit *et al.* (172) noted reduced chemotaxis and phagocytosis of peritoneal macrophages from vitamin D-deficient mice which could be corrected by vitamin D treatment. CT blocked the development of murine experimental autoimmune encephalitis (173), whereas the CT analogs OCT and 1,24(OH)₂D blocked the development of renal disease and improved survival in the MRL mouse model of autoimmunity (174, 175).

In sum, not only is vitamin D deficiency or resistance associated with subtle abnormalities in the immune system that predispose to infection, but the immunomodulatory actions of CT or its nonhypercalcemic analogs may prove to be useful agents in controlling the immune system when it goes awry. Clinical studies testing this possibility need to be performed.

D. Hypertension

1. *Role of calcium.* Considerable interest in the relationship between calcium and hypertension has developed over the past decade. Calcium is clearly necessary for the contractile response in all forms of muscle including the heart and vascular smooth muscle. Calcium channel blockers are effective means of controlling hypertension, presumably by limiting the influx of calcium into the cells responsible for maintenance of vascular tone. Thus, increased serum calcium or hormones such as PTH and CT might be expected to lead to hypertension by raising intracellular calcium concentration in the vascular smooth muscle. Consistent with this view is the observation by Erne *et al.* (176) that the intracellular free calcium concentration (Cai) of platelets correlates positively with the degree of hypertension, the positive correlation between serum calcium and blood pressure observed by Kesteloot and Geboers in 9000 young males (177), the association between hyperparathyroidism and hypertension (178, 179) and between CT levels and blood pressure (180), the increase in blood pressure induced by prolonged PTH infusions (181) (although acute PTH infusions are vasodilatory (182, 183), and the increase in blood pressure and peripheral vascular resistance seen acutely after calcium infusion (184-188). Calcium channel blockers used in the treatment of hypertension reduced platelet Cai (176) and prevented the acute increase in blood pressure during calcium infusion (185).

Although the link between calcium and hypertension seems clear, McCarron and Morris (189) have argued that it is calcium deficiency rather than excess that leads

to essential hypertension. A large number of epidemiological studies support the claim that dietary calcium deficiency and reduced serum calcium levels are associated with hypertension (reviewed in Ref. 189), and dietary calcium supplementation exerts a modest blood pressure-lowering effect in some patients especially those with the highest blood pressure and lowest serum calcium level (190-194). Renal calcium wasting has been described in patients with essential hypertension (195, 196), as has decreased intestinal calcium absorption by some (197, 198), but not all (199), groups evaluating the spontaneously hypertensive rat. Resnick and his colleagues have stratified their hypertensive patients according to renin levels. They demonstrated that the low renin group is more likely to have low serum calcium, elevated PTH and CT levels (200), and respond best to calcium supplementation (201) or calcium channel blockers (202).

2. *Role of vitamin D.* Thus, an apparent paradox exists between the presumed increase in Cai in the vascular smooth muscle and the presumed deficiency of calcium and lower serum calcium levels in hypertensive patients. In the study (176) demonstrating the increased platelet Cai in hypertensive patients, the authors also noted decreased serum ionized calcium levels, confirming this apparent paradox. Therefore, altered calcium handling by the cell has been implicated in the pathogenesis of hypertension (189), possibly at the level of reduced CaATPase activity (203), leading to increased Cai but decreased serum calcium due to decreased intestinal calcium absorption and renal calcium wasting (189). Calcium flux is regulated by CT in a number of tissues. VDR have been demonstrated in both heart (204) and vascular smooth muscle (205) cells. CT stimulates calcium flux at least in heart cells (206). Vitamin D deficiency is associated with increased contractility of both the heart and vascular smooth muscle (207, 208) suggesting increased Cai despite reduced serum calcium levels. Consistent with these findings are the observations by Baksi (209) that rats raised on either a vitamin D-deficient or calcium-deficient diet have elevated blood pressures compared to controls on normal diets. At least one clinical study has suggested that 1 α OHD could be used to treat hypertension (210).

Although these studies are provocative and suggest a role for CT and/or its analogs in the management of hypertension, considerable uncertainty remains regarding this potential application. However, if the apparent abnormality in calcium handling by cells (in particular, vascular smooth muscle cells) in hypertensive subjects is confirmed and shown to be corrected by CT, clinical trials with CT or its analogs could be initiated.

E. Diabetes mellitus

1. Calcitriol regulation of insulin secretion. The discovery that the vitamin D-dependent calcium binding protein (211–213) and the VDR (214–216) are found in the pancreas was paralleled by the discovery that vitamin D deficiency resulted in decreased insulin secretion in response to glucose or arginine (217). This abnormality could be corrected with CT. Since calcium is required for insulin secretion and since insulin secretion is blunted by starvation, the actual mechanism by which CT exerts its effects is not completely clear. Vitamin D repletion of an erstwhile vitamin D-deficient animal leads to improved nutrition and increased serum calcium levels, both of which could result in enhanced insulin secretion. The studies intended to resolve this issue have shown that pair feeding vitamin D-replete animals with vitamin D-deficient animals blunts, if not abolishes, the ability of CT to restore normal responsiveness of the islet to glucose (218–220). Thus, nutrition plays an important role, although it does not explain all the effects of CT. Raising the serum calcium level of the vitamin D-deficient rat to normal does not restore normal responsiveness of the islet to glucose (218–220). However, such dietary manipulation results in profound hypophosphatemia (219) which could itself blunt insulin secretion. Nevertheless, studies by Tanaka *et al.* (221) and Ozono *et al.* (222) demonstrated that normalization of serum calcium by diet partially corrected the defect in insulin secretion equivalent to that by CT when the rise in serum calcium was prevented by a low calcium diet. In their hands the combination of CT and calcium repletion led to the greatest degree of insulin secretion. This interaction between calcium and CT may explain the findings of Hochberg *et al.* (223) who studied six children with VDDR II. Of the five tested for insulin secretion during a glucose tolerance test, the three with the most blunted response had the lowest serum calcium levels. *In vitro* incubation of islets from vitamin D-deficient rats with CT or inclusion of CT in the islet perfusate did not restore normal insulin secretion (218). However, the response to CT *in vivo* can be seen as early as 3 h after administration (224), which is before a substantial increase in either food intake or serum calcium level. In sum, the data support a direct role of CT in promoting insulin secretion in addition to an indirect role through changes in nutrition and serum minerals.

2. Insulin regulation of CT production. Compounding the possibility that insulin secretion requires adequate levels of CT are the observations that acute insulin deficiency leads to decreased CT production (225, 226). Thus, insulin secretion and CT levels could fall rapidly and in parallel before the diabetes is recognized and treated. This may account for the observation that bone loss in

insulin-dependent diabetics occurs early, and then stabilizes unless the diabetes is poorly controlled (227–232). In BB diabetic rats, reduced CT levels are associated with decreased DBP levels (233) such that the calculated "free" CT levels are normal or even elevated. Despite this, intestinal calcium absorption, intestinal calcium binding protein, and VDR are reduced as one would find in vitamin D deficiency. Whether this estimation of the biologically available CT accurately reflects the *in vivo* situation in these animals is not clear, but if so would suggest that vitamin D resistance might also complicate the diabetic state. Observations in other experimental animal models of diabetes mellitus have also shown lower CT levels (234, 235), acute decrements in intestinal calcium transport (236), and decreased intestinal calcium binding protein (237) as well as decrements in bone formation (238).

3. Diabetes and metabolic bone disease. The clinical implications of these animal studies are not clear. Diabetes mellitus is not a well known complication of vitamin D deficiency. As mentioned previously, hypocalcemic patients with VDDR II may have a subtle defect in insulin secretion (223), but diabetes mellitus in this group has not been described. Patients with uremia (which results in decreased CT levels) are known to have abnormal carbohydrate metabolism. Mak (239) demonstrated an improvement in insulin secretion in a group of uremic subjects given CT 2 h before the glucose challenge. Conversely, reduced CT levels have been described in some (240, 241), but not all (242), studies of diabetics. The pregnant diabetic and her fetus may be at greatest risk of reduced CT levels (240), and hypocalcemia in the infant of the diabetic mother is common (242). The impact of altered vitamin D metabolism in the nonpregnant diabetic is more difficult to ascertain. Intestinal calcium absorption appears to be normal (242–244). Decreased bone density in both adult and juvenile onset diabetics has been reported by a number of groups (227–232), and the risk of fractures may be increased in this patient group (245, 246). Thus, with our current state of knowledge it is not clear that CT or its analogs has a major role to play in the management of diabetes mellitus. However, the possibility that CT or one of its analogs could enhance insulin secretion in the type 2 diabetic or prevent the loss of bone at the onset of type 1 diabetes mellitus needs to be considered.

F. Psoriasis

1. Targets for CT action. Psoriasis may be among the first of the new clinical applications for CT and its analogs to gain widespread acceptance. Psoriasis involves both an inflammatory component with infiltration of neutrophils and T lymphocytes into the dermis and a

hyperproliferative component of the epidermis with poorly differentiating keratinocytes. Both processes represent potential targets for CT. As discussed under "immune function" CT inhibits the elaboration of cytokines from activated lymphocytes and blocks their proliferation. Thus, this component of the inflammatory process seen in psoriasis may be blocked by CT. As we (247) have recently reviewed in depth, CT is also a potent modulator of keratinocyte proliferation and differentiation. Keratinocytes make CT (248, 249), contain VDR (250-252), and respond to CT with a decrease in proliferation and an increase in differentiation (251, 253, 254). As such, the hyperproliferative response of the epidermis in psoriasis should also be amenable to treatment with CT.

2. Clinical studies. Realization of the potential usefulness of CT in the treatment of psoriasis emanated from a case report in which a woman being treated for osteoporosis with 1 α OHD showed clearing of her psoriasis (255). This report was followed by a number of small open clinical trials with either 1 α OHD, 1,24(OH)₂D (comparable to CT in *in vitro* potency), or CT given orally or applied topically (256-258). Although not carefully established, it appears that the response is dose dependent (259, 260). Oral administration tends to increased serum and urine calcium levels limiting the amount of CT that can be safely given at least by this route. Data regarding the topical administration of CT suggest that higher concentrations can be administered by this route (261), but dose limitations have not been established. Because of the potential toxicity related to the use of CT, studies began with the nonhypercalcemic analog, calcipotriol (MC903). In a dose-response study Kragballe (262) used up to 100 μ g calcipotriol/g vehicle without detecting a change in serum calcium levels. This dose has been extended to 1.2 mg/g vehicle without evident toxicity (263), although maximal effectiveness was achieved at 50 μ g/g in the Kragballe study (262). These doses are 2 or 3 orders of magnitude higher than the preparations of CT, 1,24(OH)₂D, or 1 α OHD used by others (256, 258, 260, 261). Using the 50 μ g/g dose in a large multicenter study with 345 patients, Kragballe *et al.* (264) demonstrated that calcipotriol was equivalent to, if not better than, betamethasone in the treatment of psoriasis. Over 80% of the patients had marked improvement or clearing of their lesions with calcipotriol. No changes in serum calcium were reported; the only side effect noted in a significant number of subjects was burning or itching at the site of application.

These results are quite promising. However, the long term safety of the nonhypercalcemic analogs has not been established. These drugs could inhibit the normal renal production of CT and so produce a situation in

which the "classic" actions of CT are reduced (*i.e.* decreased intestinal calcium absorption). Patients with psoriasis are likely to require lifelong treatment with these CT analogs. Whether their use in high concentrations will lead to osteoporosis (reduced intestinal calcium absorption but enhanced osteoclast differentiation) or other disorders in the bone mineral homeostatic system remains for future investigation. Administering the drugs topically is likely to lead to fewer complications because the keratinocyte actively metabolizes CT (248) and is likely to do the same to the analogs thus limiting systemic exposure. Nevertheless, studies are required to investigate the long term effects of these analogs on bone mineral homeostasis before their use becomes widespread.

IV. Conclusions

An exciting new era has developed in the vitamin D field with the discovery of new target tissues, mechanisms of action, and selective analogs. In this review I have discussed the structure-function relationships of both naturally produced metabolites as well as synthetic analogs. The list was chosen not to be complete but to be illustrative. In the next few years many more analogs are likely to be available at least for research purposes. Of the available analogs, those that do not raise serum or urine calcium may be the most useful when applied to the new indications.

With the discoveries that the VDR is found in a wide (but not universal) range of tissues and that CT influences those tissues in a variety of ways comes the potential to use CT therapeutically in novel ways. The ability of CT to inhibit PTH secretion has already led to its use in the management of secondary hyperparathyroidism. Conceivably, primary hyperparathyroidism could also be treated with a nonhypercalcemic analog. The ability of CT to stimulate insulin secretion suggests the possibility that type 2 diabetics might in the future be benefitted by a CT analog. The potential for developing analogs with selective effects on osteoblast and osteoclast differentiation and function could lead to more effective use of these drugs in the management of osteoporosis and osteopetrosis. Because of the antiproliferative and pro-differentiating effect of CT on a number of cell lines including malignant cell lines, the nonhypercalcemic analogs offer an approach to the management of malignancy of those tumors that contain a VDR. Immunological disorders, including chronic inflammation, might also be managed by the CT analogs because of their potent immunomodulatory properties. Conceivably, analogs will be developed that will be selective for the different immune functions which they alter. As the abnormal cellular handling of calcium in patients with

essential hypertension becomes better defined, a role may emerge for CT or one of its analogs to modulate this process in a way that could be useful for treating this common condition. The ability of CT to modulate both the immune function and the differentiation of epidermal cells has already led to substantial success in the treatment of psoriasis.

As we enter this new era it is important to bear in mind that the analogs might be a two edged sword. By interfering with normal vitamin D metabolism and mechanisms of action of the natural metabolites, the use of high doses of the analogs could alter the bone mineral homeostatic system in a deleterious way. Such effects may not appear in short term studies when only serum and urine calcium levels are examined. These effects may be of little consequence in the management of life-threatening illnesses such as cancer but are of greater concern in the lifelong treatment of a disease such as psoriasis.

References

1. Norman AW, Koeffler HP, Bishop JE, Collins ED, Sergeev I, Zhou L-X, Nemere I, Zhou J, Henry HL, Okamura WH 1991 Structure-function relationships in the vitamin D endocrine system for 1,25(OH)₂D₃ analogs. In: Norman AW, Bouillon R, Thomasset M (eds) Vitamin D: Gene Regulation, Structure-Function Analysis and Clinical Application. Walter de Gruyter, New York, vol 1:146
2. Walters MR 1992 Newly identified actions of the vitamin D endocrine system. *Endocr Rev* 13:600
3. Procsal DA, Okamura WH, Norman AW 1976 Vitamin D, its metabolites and analogs: a review of the structural requirements for biological activity. *Am J Clin Nutr* 29:1271
4. DeLuca HF, Schnoes HK 1976 Metabolism and mechanism of action of vitamin D. *Annu Rev Biochem* 45:631
5. Haddad Jr JG 1979 Transport of vitamin D metabolites. *Clin Orthop* 142:249
6. Okamura WH, Mitra MN, Procsal DA, Norman AW 1975 Studies on vitamin D and its analogs. VIII. 3-Deoxy-1 alpha, 25-dihydroxyvitamin D₃, a potent new analog of 1 alpha, 25-(OH)₂-D₃. *Biochem Biophys Res Commun* 65:24
7. Halliwell RB, DeLuca HF 1972 Metabolites of dihydrotachysterol, in target tissues. *J Biol Chem* 247:91
8. Procsal DA, Henry HL, Friedlander EJ, Norman AW 1977 Studies on the mode of action of calciferol. *Arch Biochem Biophys* 179:229
9. Kream BE, Jose MJL, DeLuca HF 1977 The chick intestinal cytosol binding protein for 1,25 dihydroxyvitamin D₃: a study of analog binding. *Arch Biochem Biophys* 179:462
10. Eisman JA, DeLuca HF 1977 Intestinal 1,25-dihydroxyvitamin D₃ binding protein: specificity of binding. *Steroids* 30:245
11. Norman AW, Roth J, Orci L 1982 The vitamin D endocrine system: steroid metabolism, hormone receptors, and biological response (calcium binding proteins). *Endocr Rev* 3:331
12. Lam H-Y, Schnoes HK, DeLuca HF 1975 Synthesis and biological activity of 25, 26-dihydroxycholecalciferol. *Steroids* 25:247
13. Bouillon R, Allewaert K, Xiang DZ, Tan BK, Baelen HV 1991 Vitamin D analogs with low affinity for the vitamin D binding protein: enhanced *in vitro* and decreased *in vivo* activity. *J Bone Miner Res* 6:1051
14. Deleted in proof
15. Tanaka H, Hruska KA, Seino Y, Malone JD, Nishii Y, Teitelbaum SL 1991 Disassociation of the macrophage-maturational effects of vitamin D from respiratory burst priming. *J Biol Chem* 266:10888
16. Dusso AS, Negrea L, Gunawardhana S, Lopez-Hilker S, Finch J, Mori T, Nishii Y, Slatopolsky E, Brown AJ 1991 On the mechanisms for the selective action of vitamin D analogs. *Endocrinology* 128:1687
17. Farach-Carson MC, Sergeev I, Norman AW 1991 Nongenomic actions of 1,25-dihydroxyvitamin D₃ in rat osteosarcoma cells: structure-function studies using ligand analogs. *Endocrinology* 129:1876
18. Pols HAP, Birkenhager JC, Schilt JP, Bos MP, van Leeuwen JPTM 1991 The effects of MC903 on 1,25-(OH)₂D₃ receptor binding, 24-hydroxylase activity and *in vitro* bone resorption. *Bone Miner* 14:103
19. Pernalete N, Mori T, Nishii Y, Slatopolsky E, Brown AJ 1991 The activity of 22-oxacalcitriol in osteoblast-like (ROS 17/2.8) cells. *Endocrinology* 129:778
20. Evans DB, Thavarajah M, Binderup L, Kanis JA 1991 Actions of calcipotriol (MC 903), a novel vitamin D₃ analog, on human bone-derived cells: comparison with 1,25-dihydroxyvitamin D₃. *J Bone Miner Res* 6:1307
21. Valaja T, Mahonen A, Pirskanen A, Maenpaa PH 1990 Affinity of 22-oxa-1,25(OH)₂D₃ for 1,25-dihydroxyvitamin D receptor and its effects on the synthesis of osteocalcin in human osteosarcoma cells. *Biochem Biophys Res Commun* 169:629
22. Paulson SK, Perlman K, DeLuca HF, Stern PH 1990 24- and 26-homo-1,25-dihydroxyvitamin D₃ analogs: potencies on *in vitro* bone resorption differ from those reported for cell differentiation. *J Bone Miner Res* 5:201
23. Perlman K, Kutner A, Prahl J, Smith C, Inaba M, DeLuca HF 1990 24-Homologated 1,25-dihydroxyvitamin D₃ compounds: separation of calcium and cell differentiation activities. *Biochemistry* 29:190
24. Norman AW, Zhou JY, Henry HL, Uskokovic MR, Koeffler HP 1990 Structure-function studies on analogues of 1 α ,25-dihydroxyvitamin D₃: differential effects on leukemic cell growth, differentiation, and intestinal calcium absorption. *Cancer Res* 50:6857
25. Marie PJ, Connes D, Hott M, Miravet L 1990 Comparative effects of a novel vitamin D analogue MC-903 and 1,25-dihydroxyvitamin D₃ on alkaline phosphatase activity, osteocalcin and DNA synthesis by human osteoblastic cells in culture. *Bone* 11:171
26. Zhou JY, Norman AW, Lubbert M, Collins ED, Uskokovic MR, Koeffler HP 1989 Novel vitamin D analogs that modulate leukemic cell growth and differentiation with little effect on either intestinal calcium absorption or bone mobilization. *Blood* 74:82
27. Binderup L, Bramm E 1988 Effects of a novel vitamin D analogue MC 903 on cell proliferation and differentiation *in vitro* and on calcium metabolism *in vivo*. *Biochem Pharmacol* 37:889
28. Ostrem VK, Tanaka Y, Prahl J, DeLuca HF, Ikekawa N 1987 24- and 26-Homo-1,25-dihydroxyvitamin D₃: preferential activity inducing differentiation of human leukemia cells HL-60 *in vitro*. *Proc Natl Acad Sci USA* 84:2610
29. Abe J, Takita Y, Nakano T, Miyaura C, Suda T, Nishii Y 1989 A synthetic analogue of vitamin D₃, 22-oxa-1 α , 25-dihydroxyvitamin D₃, is a potent modulator of *in vitro* immunoregulating activity without inducing hypercalcemia in mice. *Endocrinology* 124:2645
30. Brown AJ, Ritter CR, Finch JL, Morrissey J, Martin KJ, Murayama E, Nishii Y, Slatopolsky E 1989 The noncalcemic analogue of vitamin D, 22-oxacalcitriol, suppresses parathyroid hormone synthesis and secretion. *J Clin Invest* 84:728
31. Abe J, Morikawa M, Miyamoto K, Kaiho S, Fukushima M, Miyaura C, Abe E, Suda T, Nishii Y 1987 Synthetic analogues of vitamin D₃ with an oxygen atom in the side chain skeleton. *FEBS Lett* 226:58
32. Brettle C, Calverley MJ, Binderup L 1991 Synthesis and biological activity of 1 α -hydroxylated vitamin D₃ analogues with hydroxylated side chains, multi-homologated in the 24- or 24,26,27-positions. In: Norman AW, Bouillon R, Thomasset M (eds) Vitamin D: Gene Regulation Structure-Function Analysis and Clinical Application. Walter de Gruyter, New York, vol 1:159
33. Abe J, Morikawa M, Takita Y, Miyamoto K, Kaiho S, Fukushima M, Miyaura C, Abe E, Suda T, Nishii Y 1988 1 α , 25-Dihydroxy-22-oxavitamin D₃: a new synthetic analogue of vitamin D₃ having potent differentiation-inducing activity without inducing hypercalcemia *in vivo* and *in vitro*. In: Norman AW, Schaefer K, Grigoleit H-G, Herrath DV (eds) Vitamin D: Molecular, Cellular

- and Clinical Endocrinology. Walter de Gruyter, New York, vol 1:310
34. Ostrem VK, Lau WF, Lee SH, Perlman K, Prahl J, DeLuca HF 1987 Induction of monocytic differentiation of HL-60 cells by 1,25-dihydroxyvitamin D analogs. *J Biol Chem* 262:14164
 35. Kabakoff BD, Kendrick NC, Faber D, DeLuca HF, Yamada S, Takayama H 1982 Determination of biological activity of 24,24-difluoro-1,25-dihydroxyvitamin D₃ in the chick using a new method for assessing intestinal calcium uptake. *Arch Biochem Biophys* 215:582
 36. Okamoto S, Tanaka Y, DeLuca HF, Kobayashi Y, Ikekawa N 1983 Biological activity of 24,24-difluoro-1,25-dihydroxyvitamin D₃. *Am J Physiol* 244:E159
 37. Shiina Y, Abe E, Miyaura C, Tanaka H, Yamada S, Ohmori M, Nakayama K, Takayama H, Matsunaga I, Nishii Y, DeLuca HF, Suda T 1983 Biological activity of 24,24-difluoro-1 alpha 25-dihydroxyvitamin D₃, and 1 alpha, 25-dihydroxyvitamin D3-26,23-lactone in inducing differentiation of human myeloid leukemia cells. *Arch Biochem Biophys* 220:90
 38. Kragballe K, Wildfang IL 1990 Calcipotriol (MC 903), a novel vitamin D₃ analogue stimulates terminal differentiation and inhibits proliferation of cultured human keratinocytes. *Arch Dermatol Res* 282:164
 39. Uskokovic MR, Baggolini E, Shiuey S-J, Iacobelli J, Hennessy B, Kiegiel J, Danielski AR, Pizzolato G, Courtney LF, Horst RL 1991 The 16-ene analogs of 1,25-dihydroxycholecalciferol. Synthesis and biological activity. In: Norman AW, Bouillon R, Thomasset M (eds) Vitamin D. Gene Regulation, Structure-Function Analysis and Clinical Application. Walter de Gruyter, New York, vol 1:139
 40. Nishii Y, Abe J, Sato K, Kobayashi T, Okano T, Tsugawa N, Slatopolsky E, Brown AJ, Dusso A, Raisz LG 1991 Characteristic of two novel vitamin D₃ analogues; 22-oxa-1 α , 25-dihydroxyvitamin D₃ [OCT] and 2 β -³(3-hydroxypropoxy)-1 α , 25-dihydroxyvitamin D₃ [ED-71]. In: Norman AW, Bouillon R, Thomasset M (eds) Vitamin D. Gene Regulation, Structure-Function Analysis and Clinical Application. Walter de Gruyter, New York, vol 1:289
 41. Bikle D, Gee E 1988 Free and not total 1,25-dihydroxyvitamin D regulates 25 hydroxyvitamin D metabolism by keratinocytes. *Endocrinology* 124:649
 42. Bikle D, Zolock DT, Morrissey RL, Herman RH 1978 Independence of 1,25-dihydroxyvitamin D₃ mediated calcium transport from *de novo* RNA and protein synthesis. *J Biol Chem* 253:484
 43. Wasserman RH, Brindak ME, Meyer SA, Fullmer CS 1982 Evidence for multiple effects of vitamin D₃ on calcium absorption: response of rachitic chicks, with or without partial vitamin D₃ repletion, to 1,25-dihydroxyvitamin D₃. *Proc Natl Acad Sci USA* 79:7939
 44. Silver J, Russell J, Sherwood LM 1985 Regulation by vitamin D metabolites of messenger ribonucleic acid for preproparathyroid hormone gene transcription *in vivo* in the rat. *Proc Natl Acad Sci USA* 82:4270
 45. Cantley LK, Russel J, Lettieri D, Sherwood LM 1985 1,25-Dihydroxyvitamin D₃ suppresses PTH secretion from bovine parathyroid cells in tissue culture. *Endocrinology* 117:2114
 46. Bikle DD, Munson S 1985 1,25-Dihydroxyvitamin D increases calmodulin binding to specific proteins in the chick duodenal brush border membrane. *J Clin Invest* 76:2312
 47. Rasmussen H, Fontaine O, Max EE, Goodman DBP 1979 Effect of 1 α -hydroxyvitamin D₃ administration on calcium transport in chick intestine brush border membrane vesicles. *J Biol Chem* 254:2993
 48. Nemere I, Yoshimoto Y, Norman AW 1984 Calcium transport in perfused duodena from normal chicks: enhancement within fourteen minutes of exposure to 1,25-dihydroxyvitamin D₃. *Endocrinology* 115:1476
 49. Lieberherr M, Grosse B, Duchampon P, Drueke T 1989 A functional cell surface type receptor is required for the early action of 1,25-dihydroxyvitamin D₃ on the phosphoinositide metabolism in rat enterocytes. *J Biol Chem* 264:20403
 50. Baran DT, Milne ML 1986 1,25-Dihydroxyvitamin D increases hepatocyte cytosolic calcium levels: a potential regulator of vitamin D-25-hydroxylase. *J Clin Invest* 77:1622
 51. Civitelli R, Kim YS, Gunstein SL, Fujimori A, Huskey M, Avioli LV, Hruska KA 1990 Nongenomic activation of the calcium message system by vitamin D metabolites in osteoblast-like cells. *Endocrinology* 127:2253
 52. McLaughlin JA, Cantley LC, Holick MF 1990 1,25(OH)₂D₃ increased calcium and phosphatidylinositol metabolism in differentiating cultured human keratinocytes. *J Nutr Biochem* 1:81
 53. Chen TL, Hirst MA, Feldman D 1979 A receptor-like binding macromolecule for 1 α , 25-dihydroxycholecalciferol in cultured mouse bone cells. *J Biol Chem* 254:7491
 54. Raisz LG, Maina DM, Gworek SG, Dietrich JW, Canalis EM 1978 Hormonal control of bone collagen synthesis *in vitro*: inhibitory effect of 1-hydroxylated vitamin D metabolites. *Endocrinology* 29:201
 55. Wong GL, Luben RA, Cohn DV 1977 1,25-dihydroxycholecalciferol and parathormone: effects on isolated osteoclast-like and osteoblast-like cells. *Science* 197:663
 56. Rowe DW, Kream BE 1982 Regulation of collagen synthesis in fetal rat calvaria by 1,25-dihydroxyvitamin D₃. *J Biol Chem* 257:8009
 57. Canalis E 1983 Effect of hormones and growth factors on alkaline phosphatase activity and collagen synthesis in cultured rat calvariae. *Metabolism* 32:14
 58. Franceschi RT, Romano PR, Park K-Y 1986 Regulation of type I collagen synthesis by 1,25-dihydroxyvitamin D₃ in human osteosarcoma (ROS 17/2.8) osteoblastic cells. *J Bone Miner Res* 1:441
 59. Owen TA, Aronow MS, Barone LM, Bettencourt B, Stein GS, Lian JB 1991 Pleiotropic effects of vitamin D on osteoblast gene expression are related to the proliferative and differentiated state of the bone cell phenotype: dependency upon basal levels of gene expression, duration of exposure, and bone matrix competency in normal rat osteoblast cultures. *Endocrinology* 128:1496
 60. Underwood JL, DeLuca HF 1984 Vitamin D is not directly necessary for bone growth and mineralization. *Am J Physiol* 246:E493
 61. Balsam S, Garabedian M, Larchet M, Groski A, Cournot G, Tau C, Bourdeau A, Silve C, Ricour C 1986 Long-term nocturnal calcium infusions can cure rickets and promote normal mineralization in hereditary resistance to 1,25-dihydroxyvitamin D. *J Clin Invest* 77:1661
 62. Bliziotis M, Yergey AL, Nanes MS, Muenzer J, Begley MG, Vieira NE, Kher KK, Brandi ML, Marx SJ 1988 Absent intestinal response to calciferols in hereditary resistance to 1,25-dihydroxyvitamin D: documentation and effective therapy with high dose intravenous calcium infusions. *J Clin Endocrinol Metab* 66:294
 63. McSheehy PMJ, Chambers TJ 1987 1,25-dihydroxyvitamin D₃ stimulates rat osteoblastic cells to release a soluble factor that increases osteoclastic bone resorption. *J Clin Invest* 80:425
 64. Suda T, Takahashi N, Martin TJ 1992 Modulation of osteoclast differentiation. *Endocr Rev* 13:66
 65. Aloia JF, Vaswani A, Yeh JK, Ellis K, Yasumura S, Cohn SH 1988 Calcitriol in the treatment of postmenopausal osteoporosis. *Am J Med* 84:401
 66. Ott SM, Chestnut CH 1989 Calcitriol treatment is not effective in postmenopausal osteoporosis. *Ann Intern Med* 110:267
 67. Gallagher JC, Goldgar D 1990 Treatment of postmenopausal osteoporosis with high doses of synthetic calcitriol. A randomized controlled study. *Ann Intern Med* 113:649
 68. Tilyard MW, Spears GFS, Thomson J, Dovey S 1992 Treatment of postmenopausal osteoporosis with calcitriol or calcium. *N Engl J Med* 326:357
 69. Caniggia A, Nuti R, Lore F, Martini G, Turchetti V, Righi G 1990 Long-term treatment with calcitriol in postmenopausal osteoporosis. *Metabolism* 39 [Suppl 1]:43
 70. Orimo H, Shiraki M, Hayashi T, Nakamura T 1987 Reduced occurrence of vertebral crush fractures in senile osteoporosis treated with 1 α (OH)-vitamin D₃. *Bone Miner* 3:47
 71. Fujita T 1990 Studies in osteoporosis in Japan. *Metabolism* 39:39
 72. Marks SC 1987 Osteopetrosis-multiple pathways for the interruption of osteoclast functions. *Appl Pathol* 5:172
 73. Yoshida H, Hayashi SI, Kunisada T, Ogawa M, Nishikawa S,

- Okamura H, Sudo T, Shultz LD, Nishikawa SI 1990 The murine mutation osteopetrosis in the coding region of the macrophage colony stimulating factor gene. *Nature* 345:442
74. Fischer A, Griscelli C, Friedrich W, Kubanek B, Levinsky R, Morgan G, Vosser J, Wagemaker G 1986 Bone marrow transplantation for immunodeficiencies and osteopetrosis: European survey 1968-1985. *Lancet* 2:1080
 75. Cournoy G, Trubert-Thil CL, Petrovic M, Boyle A, Cormier C, Girault D, Fischer A, Garabedian M 1992 Mineral metabolism in infants with malignant osteopetrosis: heterogeneity in plasma 1,25-dihydroxyvitamin D levels and bone histology. *J Bone Miner Res* 7:1
 76. Key L, Carnes D, Cole S, Holtrop M, Bar-Shavit Z, Shapiro F, Arceci R, Steinberg J, Gundberg C, Kahn A, Teitelbaum S, Anast C 1984 Treatment of congenital osteopetrosis with high-dose calcitriol. *N Engl J Med* 310:409
 77. Slatopolsky E, Lopez-Hilker S, Delmez J, Dusso A, Brown A, Martin KJ 1990 The parathyroid-calcitriol axis in health and chronic renal failure. *Kidney Int* 29:S41
 78. Brumbaugh PF, Hughes MR, Haussler MR 1975 Cytoplasmic and nuclear binding components for 1α , 25-dihydroxyvitamin D₃ in chick parathyroid glands. *Proc Natl Acad Sci USA* 72:4871
 79. Henry HL, Norman AW 1975 Studies on the mechanism of action of calciferol. VII. Localization of 1,25-dihydroxyvitamin D₃ in chick parathyroid glands. *Biochem Biophys Res Commun* 62:781
 80. Korkor AB 1987 Reduced binding of [³H] 1,25-dihydroxyvitamin D₃ in the parathyroid glands of patients with renal failure. *N Engl J Med* 316:1573
 81. Merke J, Hugel U, Zlotkowski A, Szabo A, Bommer J, Mall G, Ritz E 1987 Diminished parathyroid 1,25-(OH)₂D₃ receptors in experimental uremia. *Kidney Int* 32:350
 82. Brown EM, Wilkison RE, Eastman RC, Pallotta J, Marynick SP 1982 Abnormal regulation of parathyroid hormone release by calcium in secondary hyperparathyroidism due to chronic renal failure. *J Clin Endocrinol Metab* 54:172
 83. Russell J, Lettieri D, Sherwood LM 1986 Suppression by 1,25-(OH)₂D₃ of transcription of the parathyroid hormone gene. *Endocrinology* 119:2864
 84. Okazaki T, Igarashi T, Kronenberg HM 1988 5'-Flanking region of the parathyroid hormone gene mediates negative regulation by 1,25-(OH)₂ vitamin D₃. *J Biol Chem* 263:2203
 85. Nemeth EF, Scarpa A 1987 Receptor-dependent mobilization of cellular Ca⁺⁺ and the regulation of hormone secretion in parathyroid cells. In: Cohn DV, Martin TJ, Meunier PJ (eds) *Calcium Regulation and Bone Metabolism*. Elsevier Science, Basel, vol 1:167
 86. Yamamoto M, Igarashi T, Muramatsu M, Fukagawa EM, Moto-kura T, Ogata E 1989 Hypocalcemia increases and hypercalcemia decreases the steady-state level of parathyroid hormone messenger RNA in the rat. *J Clin Invest* 83:1053
 87. Pitts TO, Piraino BH, Mitro R, Chen TC, Segre GV, Greenberg A, Puschett JB 1988 Hyperparathyroidism and 1,25-dihydroxyvitamin D deficiency in mild, moderate and severe renal failure. *J Clin Endocrinol Metab* 67:876
 88. Malluche HH, Faugere M-C 1990 Effects of 1,25(OH)₂D₃ administration on bone in patients with renal failure. *Kidney Int* 38:S48
 89. Coburn JW 1990 Use of oral and parenteral calcitriol in the treatment of renal osteodystrophy. *Kidney Int* 38:S54
 90. Slatopolsky E, Weerts C, Thielan J, Horst R, Harter H, Martin KJ 1984 Marked suppression of secondary hyperparathyroidism by intravenous administration of 1,25-dihydroxycholecalciferol in uremic patients. *J Clin Invest* 74:2136
 91. Andress DL, Norris KC, Coburn JW, Slatopolsky E, Sherrard DJ 1989 Intravenous calcitriol in the treatment of refractory osteitis fibrosa of chronic renal failure. *N Engl J Med* 321:274
 92. Delmez JA, Tindira C, Grooms P, Dusso A, Windus DW, Slatopolsky E 1982 Parathyroid hormone suppression by intravenous 1,25-dihydroxyvitamin D. *J Clin Invest* 83:1349
 93. Dunlay R, Rodriguez M, Felsenfeld AJ, Llach F 1989 Direct inhibitory effect of calcitriol on parathyroid function (sigmoidal curve) in dialysis. *Kidney Int* 36:1093
 94. Lyles KW, Drezner MK 1982 Parathyroid hormone effects on serum 1,25-dihydroxyvitamin D levels in patients with x-linked hypophosphatemic rickets: evidence for abnormal 25-hydroxyvitamin D-1-hydroxylase activity. *J Clin Endocrinol Metab* 64:638
 95. Insogna KL, Brodus AE, Gertner JM 1983 Impaired phosphorus conservation and 1,25-dihydroxyvitamin D generation during phosphorus depletion in familial hypophosphatemic rickets. *J Clin Invest* 71:1562
 96. Scriver CR, Reade TM, DeLuca HF, Hamstra AJ 1978 Serum 1,25-dihydroxyvitamin D levels in normal subjects and in patients with hereditary rickets or bone disease. *N Engl J Med* 299:976
 97. Delvin EE, Glorieux FH 1981 Serum 1,25-dihydroxyvitamin D concentrations in hypophosphatemic vitamin D-resistant rickets. *Calif Tissue Int* 33:173
 98. Chesney RW, Mazess RB, Rose P, Hamstra AJ, DeLuca HF, Breed AL 1983 Long-term influence of calcitriol (1,25-dihydroxyvitamin D) and supplemental phosphate in X-linked hypophosphatemic rickets. *Pediatrics* 71:559
 99. Glorieux FH, Marie PJ, Pettifor JM, Delvin EE 1980 Bone response to phosphate salts, ergocalciferol and calcitriol in hypophosphatemic vitamin D-resistant rickets. *N Engl J Med* 303:1023
 100. Patron P, Gardin J-P, Borensztein P, Prigent A, Paillard M 1989 Marked direct suppression of primary hyperparathyroidism with osteitis fibrosa cystica by intravenous administration of 1,25-dihydroxycholecalciferol. *Miner Electrolyte Metab* 15:321
 101. Garland CF, Garland FC, Gorham ED 1991 Can colon cancer incidence and death rates be reduced with calcium and vitamin D? *Am J Clin Nutr* 54:193S
 102. Garland FC, Garland CF, Gorham ED, Young JF 1990 Geographic variation in breast cancer mortality in the United States: a hypothesis involving exposure to solar radiation. *Prev Med* 19:614
 103. Garland CF, Shekelle RB, Barrett-Connor E 1985 Dietary calcium and vitamin D and risk of colorectal cancer: a 19-year old prospective study in men. *Lancet* 1:307
 104. Garland CF, Comstock GW, Garland FC, Helsing KJ, Shaw EK, Gorham ED 1989 Serum 25-dihydroxyvitamin D and colon cancer: eight-year prospective study. *Lancet* 2:1176
 105. Slatter ML, Sorenson AW, Ford MH 1988 Dietary calcium intake as a mitigating factor in colon cancer. *Am J Epidemiol* 128:504
 106. Stemmermann GN, Nomura A, Chyou PH 1990 The influence of dairy and nondairy calcium on subsite large-bowel cancer risk. *Dis Colon Rectum* 33:190
 107. Kune S, Kune GA, Watson LF 1987 Case-control study of dietary etiological factors: the Melbourne colorectal cancer study. *Nutr Cancer* 9:21
 108. Simard A, Vobecky J, Vobecky JS 1991 Vitamin D deficiency and cancer of the breast: an unprovocative ecological hypothesis. *Can J Public Health* 82:300
 109. Schwartz GG, Hulka BS 1990 Is vitamin D deficiency a risk factor for prostate cancer? (hypothesis). *Anticancer Res* 10:1307
 110. Eisman JA, Barkla DH, Tutton PJH 1987 1,25-dihydroxyvitamin D₃ suppresses the *in vivo* growth of human cancer solid tumor xenografts. *Cancer Res* 47:21
 111. Manolagas SC 1987 Vitamin D and its relevance to cancer. *Anticancer Res* 7:625
 112. Colston K, Colston MJ, Feldman D 1981 1,25-Dihydroxyvitamin D₃ and malignant melanoma: the presence of receptors and inhibition of cell growth in culture. *Endocrinology* 108:1083
 113. Frampton Rd, Omund SA, Eisman JA 1983 Inhibition of human cancer cell growth by 1,25-dihydroxyvitamin D₃ metabolites. *Cancer Res* 43:4443
 114. Abe E, Miyaura C, Sakagami H, Takeda M, Konno K, Ymazaki T 1981 Differentiation of mouse myeloid leukemia cells induced by 1α , 25-dihydroxyvitamin D₃. *Proc Natl Acad Sci USA* 78:4990
 115. Miyaura C, Abe E, Kurabayashi T, Tanaka H, Konno K, Nishii Y, Suda T 1981 1α ,25-Dihydroxyvitamin D₃ induces differentiation of human myeloid leukemia cells. *Biochem Biophys Res Commun* 102:937
 116. Dodd RC, Cohen MS, Newman SL, Gray TK 1983 Vitamin D metabolites change the phenotype of monoblastic U-937 cells. *Proc Natl Acad Sci USA* 80:7538
 117. Mangelsdorf DJ, Koefoed HP, Donaldson CA, Pike JW, Haussler MR 1984 1,25-Dihydroxyvitamin D₃-induced differentiation in a

- human promyelocytic leukemia cell line (HL-60): receptor-mediated maturation to macrophage-like cells. *J Cell Biol* 98:391
118. Olsson I, Gulberg U, Ivled I, Nilsson K 1983 Induction of differentiation of the human histiocytic lymphoma cell line U-937 by 1,25-dihydroxycholecalciferol. *Cancer Res* 43:5862
 119. Koeffler HP, Amatruda T, Ikekawa N, Kobayashi Y 1984 Induction of macrophage differentiation of human normal and myeloid stem cells by 1,25-dihydroxyvitamin D₃ and its analogues. *Cancer Res* 44:5624
 120. Amento EP, Bhalla AK, Kurnick JT, Kradin RL, Holick SA, Holick MF, Krane SM 1984 1 α ,25-Dihydroxyvitamin D₃ induces maturation of the human monocyte cell line U937, and, in association with a factor from T lymphocytes, augments production of the monokine, mononuclear cell factor. *J Clin Invest* 73:731
 121. Bikle DD, Pillai S, Gee E 1992 Squamous carcinoma cell lines produce 1,25-dihydroxyvitamin D but fail to respond to its differentiating effect. *J Invest Dermatol* 97:435
 122. Pence BC, Buddingh F 1988 Inhibition of dietary-fat promoted colon carcinogenesis in rats by supplemental calcium or vitamin D₃. *Carcinogenesis* 9:187
 123. Kawaura A, Tanida N, Sawada K 1989 Supplemental administration of 1 α -hydroxyvitamin D inhibits promotion by intrarectal instillation of lithocholic acid in N-methyl-N-nitrosourea-induced colonic tumorigenesis in rats. *Carcinogenesis* 10:647
 124. Chida K, Hashiba H, Fukushima M, Suda T, Kuroki T 1985 Inhibition of tumor promotion in mouse skin by 1,25-dihydroxyvitamin D₃. *Cancer Res* 45:5426
 125. Wood AW, Chang RL, Huang M-T, Uskokovic M, Conney AH 1983 1 α ,25-Dihydroxyvitamin D₃ inhibits phorbol ester-dependent chemical carcinogenesis in mouse skin. *Biochem Biophys Res Commun* 116:605
 126. Honma Y, Hozumi M, Abe E, Konno K, Fukushima M, Hata S, Nishii Y, DeLuca HF, Suda T 1983 1 α ,25-Dihydroxyvitamin D₃ and 1 α -hydroxyvitamin D₃ prolong survival time of mice inoculated with myeloid leukemia cells. *Proc Natl Acad Sci USA* 80:201
 127. Eisman JA, Barkla DH, Tutton PJM 1987 Suppression of *in vivo* growth of human cancer solid tumor xenografts by 1,25-dihydroxyvitamin D₃. *Cancer Res* 47:21
 128. Colston KW, Berger U, Coombes RC 1989 Possible role for vitamin D in controlling breast cancer cell proliferation. *Lancet* 1:188
 129. Koeffler HP, Hirji K, Itri L 1985 1,25-Dihydroxyvitamin D₃: *in vivo* and *in vitro* effects on human preleukemic and leukemic cells. *Cancer* 62:1399
 130. Abe J, Nakano AT, Nishii Y, Matsumoto T, Ogata E, Ikeda K 1991 A novel D₃ analog, 22-oxa-1,25-dihydroxyvitamin D₃, inhibits the growth of human breast cancer *in vitro* and *in vivo* without causing hypercalcemia. *Endocrinology* 129:832
 131. Zhou JY, Norman AW, Chen DL, Sun GW, Uskokovic M, Koeffler HP 1990 1,25-Dihydroxy-16-ene-23-yne-vitamin D₃ prolongs survival time of leukemic mice. *Proc Natl Acad Sci USA* 87:3929
 132. Breslau NA, McGuire JL, Zerwekh JE, Frankel EF, Pak CYC 1984 Hypercalcemia associated with increased serum calcitriol levels in three patients with lymphoma. *Ann Intern Med* 100:1
 133. Mudde AH, van den Berg H, Boshuis PG 1987 Ectopic production of 1,25-dihydroxyvitamin D by B-cell lymphoma as a cause of hypercalcemia. *Cancer* 59:1543
 134. Needle MA, Chandra B 1984 Hypercalcemia, Hodgkin's disease and calcitriol. *Ann Intern Med* 100:916
 135. Zaloga GP, Eil C, Medbery CA 1985 Humoral hypercalcemia in Hodgkin's disease. Association with elevated 1,25-dihydroxycholecalciferol levels and subperiosteal bone reabsorption. *Arch Intern Med* 145:155
 136. Davies M, Hayes ME, Mawer EB, Lumb GA 1985 Abnormal vitamin D metabolism in Hodgkin's lymphoma. *Lancet* 1:1186
 137. Rosenthal N, Insogna KL, Godsall JW, Smaldone L, Waldron JA, Stewart AF 1985 Elevations in circulating 1,25-dihydroxyvitamin D in three patients with lymphoma-associated hypercalcemia. *J Clin Endocrinol Metab* 60:29
 138. Schaefer K, Saupe J, Pauls A, von Herrath D 1986 Hypercalcemia and elevated serum 1,25-dihydroxyvitamin D₃ in a patient with Hodgkin's lymphoma. *Klin Wochenschr* 64:89
 139. Rieke JW, Donaldson SS, Horning SJ 1989 Hypercalcemia and vitamin D metabolism in Hodgkin's disease. *Cancer* 63:1700
 140. Mercier RJ, Thompson JM, Messerschmidt GL 1988 Recurrent hypercalcemia and elevated 1,25-dihydroxyvitamin D levels in Hodgkin's disease. *Am J Med* 84:165
 141. Lambrecht L, Boelaert J, Louwagie A, Criel A, Daniels R, Bouillat R 1986 Hypercalcemia in Hodgkin's disease. *Acta Clin Belg* 1:37
 142. Fetchick DA, Bertolini DR, Sarin PS, Weintraub ST, Mundy GR, Dunn JF 1986 Production of 1,25-dihydroxyvitamin D₃ by human T-cell lymphotropic virus-1-transformed lymphocytes. *J Clin Invest* 78:592
 143. Bhalla AK, Amento EP, Clemens TL, Holick MF, Krane SM 1988 Specific high-affinity receptors for 1,25-dihydroxyvitamin D₃ in human peripheral blood mononuclear cells: presence in monocytes and induction in T lymphocytes following activation. *J Clin Endocrinol Metab* 57:1308
 144. Provvedini DM, Tsoukas CD, Deftois LJ, Manolagas SC 1983 1,25-Dihydroxyvitamin D₃ receptors in human leukocytes. *Science* 221:181
 145. Rigby WFC, Denome S, Fanger MW 1992 Regulation of lymphokine production and human T cell activation by 1,25-dihydroxyvitamin D₃: specific inhibition at the level of messenger RNA. *J Clin Invest* 79:1659
 146. Reichel H, Koeffler HP, Tobler A, Norman AW 1987 1 α 25-Dihydroxyvitamin D₃ inhibits γ -interferon synthesis by normal human peripheral blood lymphocytes. *Proc Natl Acad Sci USA* 84:3385
 147. Tobler A, Gasson J, Reichel H, Norman AW, Koeffler HP 1987 Granulocyte-macrophage colony-stimulating factor. Sensitive and receptor-mediated regulation by 1,25-dihydroxyvitamin D₃ in human peripheral blood lymphocytes. *J Clin Invest* 79:1700
 148. Bhalla AK, Amento EP, Krane SM 1986 Differential effects of 1,25-dihydroxyvitamin D₃ on human lymphocytes and monocytes/macrophage: inhibition of interleukin 2 and augmentation of interleukin 1 production. *Cell Immunol* 98:311
 149. Manolagas SC, Hustmyer FG, Yu X 1990 Immunomodulating properties of 1,25-dihydroxyvitamin D₃. *Kidney Int* 29:S9
 150. Rook GAW, Taverne J, Leveton C, Steele J 1987 The role of gamma interferon, vitamin D₃ metabolites and tumor necrosis factor in the pathogenesis of tuberculosis. *Immunology* 62:229
 151. Zarabanda MT, Riancho JA, Amado JA, Olmos JM, Gonzalez-Macias J 1992 Effect of calcitriol on the secretion of prostaglandin E₂, interleukin 1, and tumor necrosis factor α by human monocytes. *Bone* 13:185
 152. Cohen MS, Mesler DE, Snipes RG, Gray TK 1986 Vitamin D₃ activates secretion of hydrogen peroxide by human monocytes. *J Immunol* 136:1049
 153. Polia BS, Healy AM, Amento EP, Krane SM 1986 1,25-Dihydroxyvitamin D₃ maintains adherence of human monocytes and protects them from thermal injury. *J Clin Invest* 77:1332
 154. Iho S, Takahashi T, Kura F, Sugiyana H, Hoshino T 1986 The effect 1,25-dihydroxyvitamin D₃ on *in vitro* immunoglobulin production by B cells. *J Immunol* 136:4427
 155. Provvedini DM, Tsoukas CD, Deftois LJ, Manolagas SC 1986 1,25-Dihydroxyvitamin D₃ binding macromolecules in human B lymphocytes: effects on immunoglobulin production. *J Immunol* 136:2734
 156. Adams JS, Gacad MA 1985 Characterization of 1 α -hydroxylation of vitamin D₃ sterols by cultured alveolar macrophages from patients with sarcoidosis. *J Exp Med* 161:755
 157. Koeffler HP, Reichel H, Bishop JE, Norman AW 1985 Gamma-Interferon stimulates production of 1,25-dihydroxyvitamin. *Biochem Biophys Res Commun* 127:596
 158. Mason RS, Frankel T, Chan YL, Lissner D, Posen S 1984 Vitamin D conversion by sarcoid lymph node homogenate. *Ann Intern Med* 100:59
 159. Adams JS, Sharma OP, Gacad MA, Singer FR 1983 Metabolism of 25-hydroxyvitamin D₃ by cultured pulmonary alveolar macrophages in sarcoidosis. *J Clin Invest* 72:1856
 160. Cadarrel J, Hance AJ, Milleron B, Paillard F, Akoun GM, Garabedian M 1988 Production of 1,25(OH)₂D₃ by cells recovered by

- bronchoalveolar lavage and the role of this metabolite in calcium homeostasis. *Am Rev Respir Dis* 138:984
161. Barnes PF, Modlin RL, Bikle DD, Adams JS 1989 Transpleural gradient of 1,25-dihydroxyvitamin D in tuberculous pleuritis. *J Clin Invest* 83:1527
 162. Stroder J, Kasal P 1970 Evaluation of phagocytosis in rickets. *Acta Paediatr Scand* 59:288
 163. Lorente F, Fontan G, Jara P, Casas C, Garcia-Rodriguez MC, Ojeda JA 1976 Defective neutrophil motility in hypovitaminosis D rickets. *Acta Paediatr Scand* 65:695
 164. Davies PDO 1985 A possible link between vitamin D deficiency and impaired host defence to *Mycobacterium tuberculosis*. *Tubercle* 66:301
 165. Toss G, Symrung T 1983 Delayed hypersensitivity response and vitamin D deficiency. *Int J Vitam Nutr Res* 53:27
 166. Tabata T, Suzuki R, Kikunami K, Matsushita Y, Inoue T, Okamoto T, Miki T, Nishizawa Y, Morii H 1986 The effect of 1 α -hydroxyvitamin D₃ on cell-mediated immunity in hemodialyzed patients. *J Clin Endocrinol Metab* 63:1218
 167. Walka MM, Daumling S, Hadorn H-B, Kruse K, Belohradsky BH 1991 Vitamin D dependent rickets type II with myelofibrosis and immune dysfunction. *Eur J Pediatr* 150:665
 168. Etzion A, Hochberg Z, Pollak S, Meshulam T, Zakut V, Tzeboval E, Keisari Y, Aviram I, Spirer Z, Benderly A, Weisman Y 1989 Defective leukocyte fungicidal activity in end-organ resistance to 1,25-dihydroxyvitamin D. *Pediatr Res* 25:276
 169. Kitajima I, Maruyama I, Matsubara H, Osame M, Igata A 1989 Immune dysfunction in hypophosphatemic vitamin D-resistant rickets: immunoregulatory reaction of 1 α (OH) vitamin D₃. *Clin Immunol Immunopathol* 53:24
 170. Huckins D, Felson DT, Holick M 1990 Treatment of psoriatic arthritis with oral 1,25 dihydroxyvitamin D₃: a pilot study. *Arthritis Rheum* 33:1723
 171. Stroder J, Schneider J 1975 Immunity in vitamin D deficient rickets. In: Norman AW, Schaefer K, Herrath DV, Ritz E (eds) *Vitamin D and Problems Related to Uremic Bone Disease*. Walter de Gruyter, New York, vol 1:675
 172. Bar-Shavit Z, Noff D, Edelstein S, Meyer M, Shibolet S, Goldman R 1981 1,25-Dihydroxyvitamin D₃ and the regulation of macrophage function. *Calcif Tissue Int* 33:673
 173. Lemire JM, Archer DC 1991 1,25-Dihydroxyvitamin D₃ prevents the *in vivo* induction of murine experimental autoimmune encephalomyelitis. *J Clin Invest* 87:1103
 174. Koizumi T, Nakao Y, Matsui T, Nakagawa T, Matsuda S, Komoriya K, Kanai Y, Fujita T 1985 Effects of corticosteroid and 1,24R-dihydroxy-vitamin D₃ administration on lymphoproliferation and autoimmune disease in MRL/MP-1pr/1pr mice. *Int Arch Allergy Appl Immunol* 77:396
 175. Abe J, Takita Y, Nakano T, Miura C, Suda T, Nishii Y 1989 22-OXA-1 α ,25-Dihydroxyvitamin D₃: a new synthetic analogue of vitamin D₃ having a potent immunoregulatory activity without inducing hypercalcemia in mice. *J Bone Miner Res* 4:S260
 176. Erne P, Bolli P, Burgisser E, Buhler FR 1984 Correlation of platelet calcium with blood pressure, effect of antihypertensive therapy. *N Engl J Med* 310:1084
 177. Kesteloot H, Geboers J 1982 Calcium and blood pressure. *Lancet* 1:813
 178. Rosenthal FD, Roy S 1972 Hypertension of hyperparathyroidism. *Br Med J* 4:396
 179. Hellstrom J, Birke G, Edvall CA 1958 Hypertension in hyperparathyroidism. *Br J Urol* 11:369
 180. Sowers MFR, Wallace RB, Hollis BW, Lemke JH 1988 Relationships between 1,25-dihydroxyvitamin D and blood pressure in a geographically defined population. *Am J Clin Nutr* 48:1053
 181. Hulter HN, Melby JC, Peterson JC, Cooke CR 1986 Chronic continuous PTH infusion results in hypertension in normal subjects. *J Clin Hypertens* 2:360
 182. Collip JB, Clark EP 1925 Further studies on the physiological action of parathyroid hormone. *J Biol Chem* 64:485
 183. Pang PKT, Tenner Jr TE, Yee JA 1980 Hypotensive action of parathyroid hormone preparations on rats and dogs. *Proc Natl Acad Sci USA* 77:675
 184. Tresham JJ, McGuire P, Coghlan JP, Whitworth JA, Scoggins BA 1988 The effects of calcium and vitamin D on blood pressure in conscious sheep. *Clin Exp Hypertens A10:1085*
 185. Berl T, Levi M, Ellis M, Chaimovitz C 1985 Mechanism of acute hypercalcemic hypertension in the conscious rat. *Hypertension* 7:923
 186. Bianchetti MG, Beretta-Piccoli C, Weidmann P, Link L, Boehringer K, Ferrier C, Morton JJ 1983 Calcium and blood pressure regulation in normal and hypertensive subjects. *Hypertension* 5:57
 187. Marone C, Beretta-Piccoli C, Weidmann P 1980 Acute hypercalcemic hypertension in man: role of haemodynamics, catecholamines and renin. *Kidney Int* 20:92
 188. Weidmann P, Massry SG, Coburn JW, Maxwell MH, Atleson J, Leeman CR 1972 Blood pressure effect of acute hypercalcemia. *Ann Intern Med* 76:741
 189. McCarron DA, Morris CD 1986 Metabolic considerations and cellular mechanism related to calcium's antihypertensive effects. *Fed Proc* 45:2734
 190. Grobbee DE, Hofman A 1986 Effect of calcium supplementation on diastolic blood pressure in young people with mild hypertension. *Lancet* 2:703
 191. Belizan JM, Villar J, Pineda O 1983 Reduction of blood pressure with calcium supplementation in young adults. *JAMA* 249:1161
 192. McCarron DA, Morris CD 1985 Blood pressure response to oral calcium in persons with mild to moderate hypertension. *Ann Intern Med* 103:825
 193. Lyle RM, Melby CL, Hyner GC, Edmondson JW, Miller JZ, Weinberger MH 1987 Blood pressure and metabolic effects of calcium supplementation in normotensive white and black males. *JAMA* 257:1772
 194. Johnson NE, Smith EL, Freudenheim JL 1985 Effects on blood pressure of calcium supplementation of women. *Am J Clin Nutr* 42:12
 195. Strazzullo P, Nunziata V, Cirillo M 1983 Abnormalities of calcium metabolism in essential hypertension. *Clin Sci* 65:137
 196. McCarron DA, Pingree PA, Rubin RJ, Gaucher SM, Molitch M, Krutzik S 1980 Enhanced parathyroid hormone function in essential hypertension: a homeostatic response to a urinary calcium leak. *Hypertension* 2:162
 197. Schedl HP, Miller DL, Pape JM, Horst RL, Wilson HD 1984 Calcium and sodium transport and vitamin D metabolism in the spontaneously hypertensive rat. *J Clin Invest* 73:980
 198. McCarron DA, Lucas PA, Lacour B, Shneidman RJ, Druke T 1985 Blood pressure development of the spontaneously hypertensive rat following concurrent manipulations of dietary Ca²⁺ and Na⁺: relation to intestinal Ca²⁺ fluxes. *J Clin Invest* 76:1147
 199. Lau K, Langman CB, Gafte U, Dudeja PK, Brasitus TA 1986 Increased calcium absorption in prehypertensive spontaneously hypertensive rat. *J Clin Invest* 78:1083
 200. Resnick LM, Muller FB, Laragh JH 1986 Calcium regulating hormones in essential hypertension: relation to plasma renin activity and sodium metabolism. *Ann Intern Med* 105:649
 201. Resnick LM, Nicholson JP, Laragh JH 1986 Calcium metabolism in essential hypertension: relationship to altered renin system activity. *Fed Proc* 45:2739
 202. Nicholson JP, Resnick LM, Laragh JH 1987 The antihypertensive effect of verapamil at extremes of dietary sodium intake. *Ann Intern Med* 107:329
 203. Postnov YV, Orlov SN, Pokudin NI 1979 Decrease in calcium binding by the red blood cell membrane in spontaneously hypertensive rats and in essential hypertension. *Pfluegers Arch* 379:191
 204. Walters MR, Wicker DC, Riggle PC 1986 1,25-Dihydroxyvitamin D receptors identified in the rat heart. *J Mol Cell Cardiol* 18:67
 205. Merke J, Hofmann W, Goldschmidt G, Ritz E 1987 Demonstration of 1,25(OH)₂ vitamin D₃ receptors and actions in vascular smooth muscle. *Calcif Tissue Int* 41:1706
 206. Walters MR, Ilenchuk TT, Claycomb WC 1987 1,25(OH)₂D₃ stimulates ⁴⁵Ca²⁺ uptake by cultured adult rat ventricular cardiac muscle cells. *J Biol Chem* 262:2536
 207. Hochhauser E, Barak J, Kushnir T, Navon G, Meyer MS, Edelstein S, Bassat MB, Vidne BA 1989 Mechanical, biochemical, and

- structural effects of vitamin D deficiency on the chick heart. *Angiology* 1:300
208. Weishaar RE, Simpson RU 1987 Vitamin D₃ and cardiovascular function in rats. *J Clin Invest* 79:1706
209. Baksi SN 1988 Altered pressor response to norepinephrine in calcium and vitamin D-deficient rats. *Clin Exp Hypertens* A10:811
210. Ljunghall S, Hvarfner A, Lind L 1987 Clinical studies of calcium metabolism in essential hypertension. *Eur Heart J* 8:37
211. Christakos S, Norman AW 1979 Studies on the mode of action of calciferol. XVII. Evidence for a specific high affinity binding protein for 1,25-dihydroxyvitamin D₃ in chick kidney and pancreas. *Biochem Biophys Res Commun* 89:56
212. Morrissey RL, Bucci TJ, Empson RN, Lufkin EG 1975 Calcium binding protein: its cellular localization in jejunum, kidney and pancreas. *Proc Soc Exp Biol Med* 149:56
213. Arnold BB, Kuttner M, Willis DM, Hitchman JW, Harrison JE, Murray TM 1975 Radioimmunoassay studies of intestinal calcium binding protein in the pig. II. The distribution of intestinal CaBP in pig tissues. *Can J Physiol Pharmacol* 53:1135
214. Christakos S, Norman AW 1981 Studies on the mode of action of calciferol. XXIX. Biochemical characterization of 1,25-dihydroxyvitamin D₃ receptors in chick pancreas and kidney cytosol. *Endocrinology* 108:140
215. Pike JW 1981 Receptors for 1,25-dihydroxyvitamin D₃ in chick pancreas: a partial physical and functional characterization. *J Steroid Biochem* 16:385
216. Clark SA, Stumpf WE, Sar M, DeLuca HF, Tanaka Y 1980 Target cells for 1,25-dihydroxyvitamin D₃ in the pancreas. *Cell Tissue Res* 209:515
217. Norman AW, Frankel BJ, Heldt AM, Grodsky GM 1980 Vitamin D deficiency inhibits pancreatic secretion of insulin. *Science* 209:823
218. Chertow BS, Sivitz WI, Baranetsky NG, Clark SA, Waite A, DeLuca HF 1983 Cellular mechanisms of insulin release: the effects of vitamin deficiency and repletion on rat insulin secretion. *Endocrinology* 113:1511
219. Cade C, Norman AW 1986 Vitamin D₃ improves impaired glucose tolerance and insulin secretion in the vitamin D-deficient rat *in vivo*. *Endocrinology* 119:84
220. Kadawaki S, Norman AW 1984 Dietary vitamin D is essential for normal insulin secretion from perfused rat pancreas. *J Clin Invest* 73:759
221. Tanaka Y, Seino Y, Ishida M, Yamaoka K, Satomura K, Yabuuchi H, Imura H 1986 Effect of 1,25-dihydroxyvitamin D₃ on insulin secretion: direct or mediated? *Endocrinology* 118:1971
222. Ozono K, Seino Y, Yano H, Yamaoka K 1990 1,25-Dihydroxyvitamin D₃ enhances the effect of refeeding on steady state preproinsulin messenger ribonucleic acid levels in rats. *Endocrinology* 126:2041
223. Hochberg Z, Borochowitz Z, Benderli A, Vardi P, Oren S, Spire Z, Heyman I, Weissman Y 1985 Does 1,25-dihydroxyvitamin D participate in the regulation of hormone release from endocrine glands? *J Clin Endocrinol Metab* 60:57
224. Cade C, Norman AW 1987 Rapid normalization/stimulation by 1,25-dihydroxyvitamin D₃ of insulin secretion and glucose tolerance in the vitamin D-deficient rat. *Endocrinology* 120:1490
225. Spencer EM, Khalil M, Tobiasen O 1980 Experimental diabetes in the rat causes an insulin reversible decrease in renal 25-dihydroxyvitamin D₃-1 α -hydroxylase activity. *Endocrinology* 107:300
226. Wongsurawat N, Armbrecht HJ, Zenser TV, Davis BB, Thomas ML, Forte LR 1983 1,25-dihydroxyvitamin D₃ and 24,25-dihydroxyvitamin D₃ production by isolated renal slices is modulated by diabetes and insulin in the rat. *Diabetes* 32:302
227. Mathiassen B, Sielsen S, Johansen JS, Hartwell D, Ditzel J, Christiansen C 1990 Long-term bone loss in insulin-dependent diabetic patients with microvascular complications. *J Diabet Complications* 4:145
228. McNair P 1988 Bone mineral metabolism in human type 1 (insulin dependent) diabetes mellitus. *Dan Med Bull* 35:109
229. Shore RM, Chesney RW, Mazess RB, Rose PG, Bargmann GJ 1981 Osteopenia in juvenile diabetes. *Calcif Tissue Int* 33:455
230. Hui SL, Epstein S, Johnston CC 1985 A prospective study of bone mass in patients with type 1 diabetes. *J Clin Endocrinol Metab* 60:74
231. Levin M, Boisseau V, Avioli L 1976 Effects of diabetes mellitus on bone mass in juvenile and adult onset diabetes. *N Engl J Med* 294:241
232. Ihida H, Seino Y, Matsukura S, Ikeda M, Yawata M, Yamahita G, Ishizuka S, Imura H 1985 Diabetic osteopenia and circulating levels of vitamin D metabolites in type 2 (noninsulin independent) diabetes. *Metabolism* 34:797
233. Nyomba BL, Verhaeghe J, Thomasset M, Lissens W, Bouillon R 1989 Bone mineral homeostasis in spontaneously diabetic BB rats. I. Abnormal vitamin D metabolism and impaired active intestinal calcium absorption. *Endocrinology* 124:565
234. Schneider LE, Schedl HP, McCain T, Haussler MR 1977 Experimental diabetes reduces circulating 1,25-dihydroxyvitamin D in the rat. *Science* 196:1452
235. Hough S, Fausto A, Sonn Y, Don JK, Birge SJ, Avioli LV 1983 Vitamin D metabolism in the chronic streptozotocin-induced diabetic. *Endocrinology* 113:790
236. Schneider LE, Schedl HP 1972 Diabetes and intestinal calcium absorption in the rat. *Am J Physiol* 223:1319
237. Schneider LE, Wilson HD, Schedl HP 1973 Intestinal calcium binding protein in the diabetic rat. *Nature* 245:327
238. Goodman WG, Hori MT 1984 Diminished bone formation in experimental diabetes. Relationship to osteoid maturation and mineralization. *Diabetes* 33:825
239. Mak RHK 1989 Insulin secretion in uremia: effect of parathyroid hormone and vitamin D metabolites. *Kidney Int* 36:S227
240. Kuoppala T 1988 Alterations in vitamin D metabolites and minerals in diabetic pregnancy. *Gynecol Obstet Invest* 25:99
241. Frazer T, White N, Hough S, Santiago J, McGee B, Bryce G, Mallon J, Avioli L 1981 Alterations in circulating vitamin D metabolites in the young insulin-dependent diabetic. *J Clin Endocrinol Metab* 93:1154
242. Heath III H, Lambert P, Service F, Arnaud S 1979 Calcium homeostasis in diabetes mellitus. *J Clin Endocrinol Metab* 49:462
243. Tsang R, Kleinman L, Sutherland J, Light I 1972 Hypocalcemia in infants of diabetic mothers. *J Pediatr* 80:384
244. Witt MF, White NH, Santiago JV, Seino Y, Avioli LV 1982 Increased calcium absorption in type I diabetic children (IDD). *Pediatr Res* 16:1129
245. Heath H, Melton JL, Chu-Pin C 1980 Diabetes mellitus and risk of skeletal fracture. *N Engl J Med* 303:567
246. Bouillon R 1991 Diabetic bone disease. *Calcif Tissue Int* 49:155
247. Bikle DD, Pillai S 1992 Vitamin D, calcium and epidermal differentiation. *Endocr Rev*, in press
248. Bikle DD, Nemani MK, Gee EA 1986 1,25-Dihydroxyvitamin D₃ production by human keratinocytes: kinetics and regulation. *J Clin Invest* 78:557
249. Bikle DD, Nemani MK, Whitney JO, Elias PW 1986 Neonatal human foreskin keratinocytes produce D₃. *Biochem Biophys Res Commun* 25:1545
250. Stumpf WE, Sar M, Reid FA, Tanaka Y, DeLuca HF 1979 Target cells for 1,25-dihydroxyvitamin D₃ in intestinal tract, stomach, kidney, skin, pituitary and parathyroid. *Science* 206:1188
251. Hosomi J, Hosoi J, Abe E, Suda T, Kuroki T 1983 Regulation of terminal differentiation of cultured mouse epidermal cells by 1 α ,25-dihydroxyvitamin D₃. *Endocrinology* 113:1950
252. Pillai S, Bikle DD, Elias PM 1988 1,25-Dihydroxyvitamin D production and receptor binding in human keratinocytes varies with differentiation. *J Biol Chem* 263:5390
253. Smith EL, Walworth ND, Holick MF 1986 Effect of 1,25-dihydroxyvitamin D₃ on the morphologic and biochemical differentiation of cultured human epidermal keratinocytes grown in serum free conditions. *J Invest Dermatol* 86:709
254. Pillai S, Bikle DD 1991 Role of intracellular-free calcium in the cornified envelope formation of keratinocytes: differences in the mode of action of extracellular calcium and 1,25 dihydroxyvitamin D₃. *J Cell Physiol* 146:94

255. Morimoto S, Kumahara Y 1986 A patient with psoriasis cured by 1 α -hydroxyvitamin D₃. *J Invest Dermatol* 86:709
256. Morimoto S, Yoshikawa K, Kozuka T 1986 An open study of vitamin D₃ treatment in psoriasis. *Br J Dermatol* 115:431
257. Morimoto S, Yoshikawa K, Kozuka T, Kitano Y, Imanaka S, Fukuo K, Koh E, Onishi T, Kumahara Y 1986 Treatment of Psoriasis vulgaris by oral administration of 1 α -hydroxyvitamin D₃-open-design study. *Calcif Tissue Int* 39:209
258. Kato T, Rokugo M, Terui T, Tagami H 1986 Successful treatment of psoriasis with topical application of active vitamin D₃ analogue, 1 α ,24-dihydroxycholecalciferol. *Br J Dermatol* 115:431
259. Henderson CA, Papworth-Smith J, Cunliffe WJ, Hight AS, Shamy HK, Czarnetzki BM 1989 A double-blind, placebo-controlled trial of topical 1,25-dihydroxycholecalciferol in psoriasis. *Br J Dermatol* 121:493
260. Smith EL, Pincus SH, Donovan L, Holick MF 1988 A novel approach for the evaluation and treatment of psoriasis. or topical use of 1,25-dihydroxyvitamin D₃ can be a safe and effective therapy for psoriasis. *J Am Acad Dermatol* 19:516
261. Holick MF 1989 Will 1,25-dihydroxyvitamin D₃, MC 903, and their analogues herald a new pharmacologic era for the treatment of psoriasis. *Arch Dermatol Res* 125:1692
262. Kragballe K 1989 Treatment of psoriasis by the topical application of the novel cholecalciferol analogue calcipotriol (MC 903). *Arch Dermatol Res* 125:1647
263. Staberg B, Roed-Petersen J, Menne T 1989 Efficacy of topical treatment in psoriasis with MC 903, a new vitamin D analogue. *Acta Derm Venereol (Stockh)* 69:147
264. Kragballe K, Gjertsen BT, DeHoop D, Karlsmark T, van De Kerkhof PCM, Larko O, Tikjob G, Nieboer C, Strand A, Roed-Petersen J 1991 Double-blind, right/left comparison of calcipotriol and betamethasone valerate in treatment of psoriasis vulgaris. *Lancet* 337:193